

# **Métodos de Inteligencia Artificial en Biología Computacional Artificial Intelligence Methods in Computational Biology**

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# Computational Biology. Definition

**Bioinformatics:** Research, development, or application of computational tools and approaches for expanding the use of biological, medical, behavioral or health data, including those to acquire, store, organize, archive, analyze, or visualize such data.

**Computational Biology:** The development and application of data-analytical and theoretical methods, mathematical modeling and computational simulation techniques to the study of biological, behavioral, and social systems.

Bioinformatics applies principles of information sciences and technologies to make the vast, diverse, and complex life sciences data more understandable and useful. Computational biology uses mathematical and computational approaches to address theoretical and experimental questions in biology.

NIH Working Definitions of Bioinformatics and Computational biology, 2000:

<https://www.bisti.nih.gov/docs/CompuBioDef.pdf>

The NIH Biomedical Information Science and Technology Initiative Consortium agreed on the previous definitions of bioinformatics and computational biology recognizing that no definition could completely eliminate overlap with other activities or preclude variations in interpretation by different individuals and organizations.

# Computational Biology. Definition

Other definition:

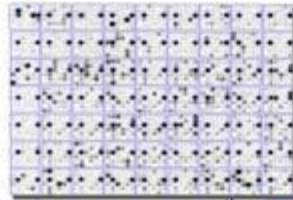
**Computational biology:** the study of biology using computational techniques. The goal is to learn new biology, knowledge about living systems. **It is about science.**

**Bioinformatics:** the creation of tools (algorithms, databases) that solve problems. The goal is to build useful tools that work on biological data. **It is about engineering.**

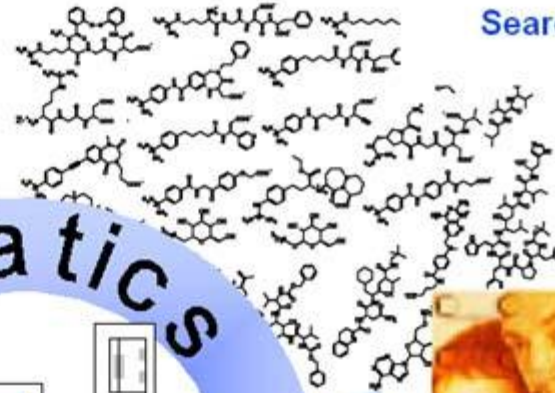
(Russ Altman, Stanford University).

# Computational Biology. Areas of research

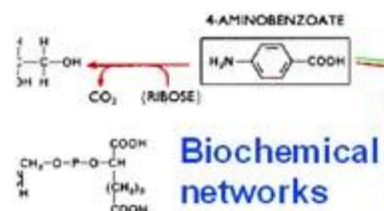
DNA chips: comparison of cell states



Search for new drugs

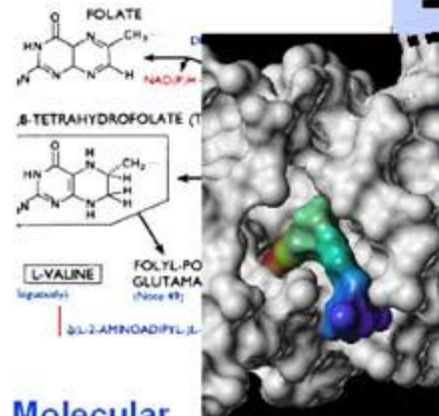


Genetic variations



Data handling, Algorithms  
Statistics, Visualisation

Optimizing therapies



Genomes

```
cacctggggccaccaccctagggtggcca
atctactcccaggagccggaggggccggag...
```

Proteins

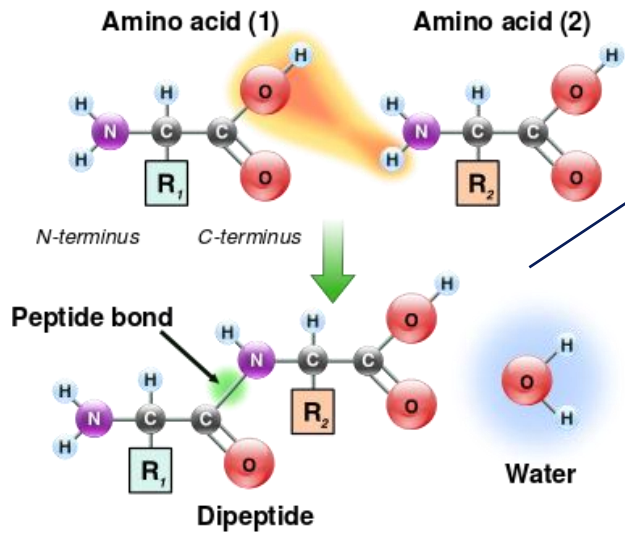
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MTNRNFRQINLLDLR VQR RVPVIHQETA
ECGLACLAMICGHFGKNIDIIYLRRKFNL...
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Sequence analysis

© Thomas Lengauer

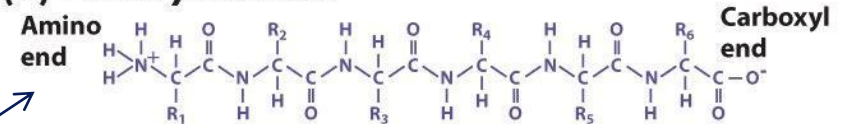
# Protein structure prediction and protein folding modeling with cellular automata

# Proteins

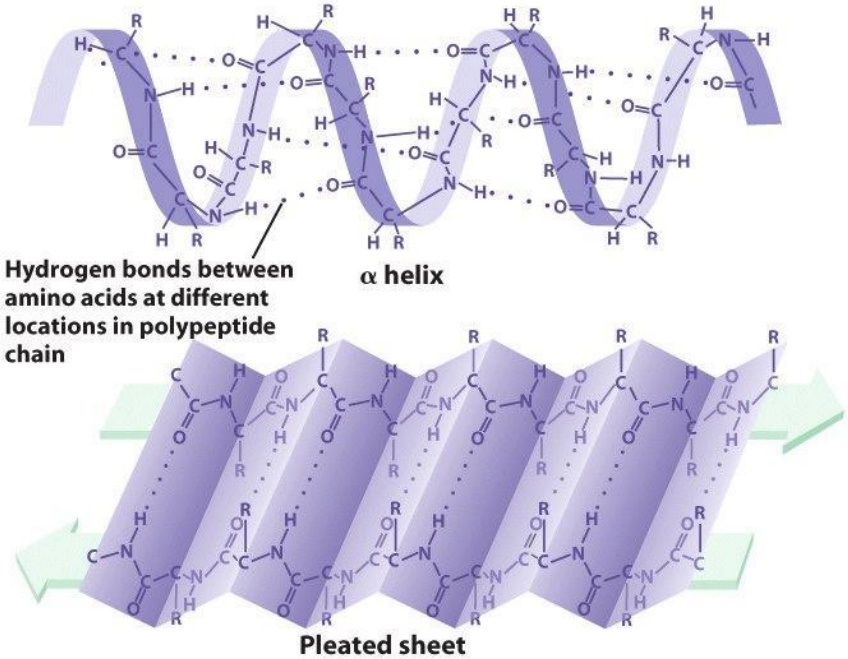


“Ab initio”  
prediction

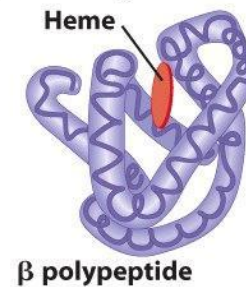
## (a) Primary structure



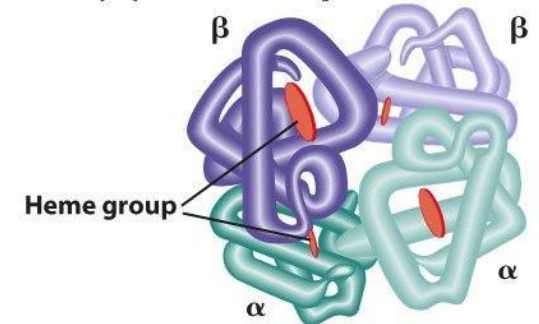
## (b) Secondary structure

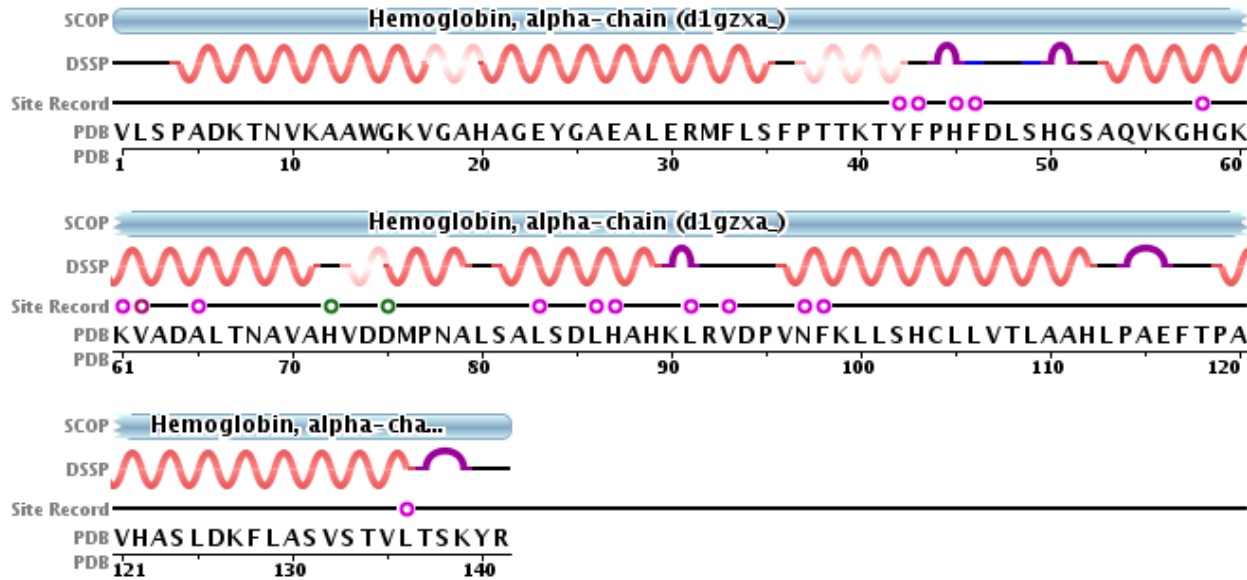


## (c) Tertiary structure



## (d) Quaternary structure





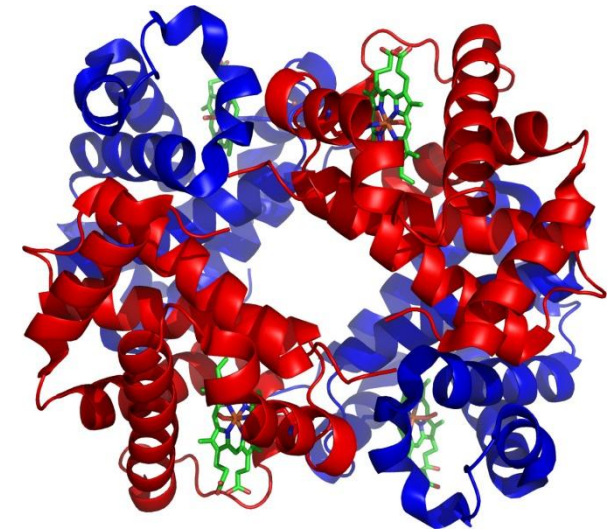
### Site Record Legend

- BINDING SITE FOR RESIDUE OXY A1143 (SOFTWARE)
- BINDING SITE FOR RESIDUE HEM B1290 (SOFTWARE)
- BINDING SITE FOR RESIDUE HEM A1142 (SOFTWARE)

### DSSP Legend

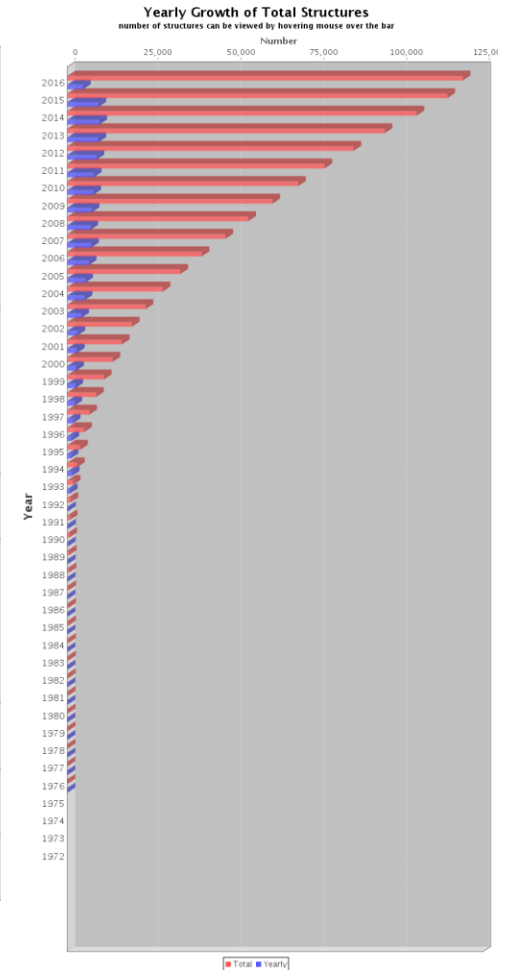
- T: turn
- empty: no secondary structure assigned
- G: 3/10-helix
- S: bend
- H: alpha helix

“Ab initio”  
prediction



# Proteins - PDB (Protein Data Bank)

Experimental Method	<u>Proteins</u>	<u>Nucleic Acids</u>	Protein/Nucleic Acid complexes	Other	Total
<u>X-ray diffraction</u>	95636	1694	4817	4	102151
<u>NMR</u>	9840	1135	231	8	11214
<u>Electron microscopy</u>	666	29	227	0	922
Hybrid	83	3	2	1	89
Other	170	4	6	13	193
<i>Total:</i>	106293	2865	5283	26	<b>114569</b>



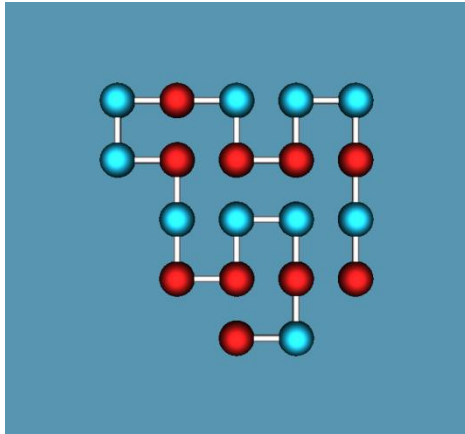


# Proteins.

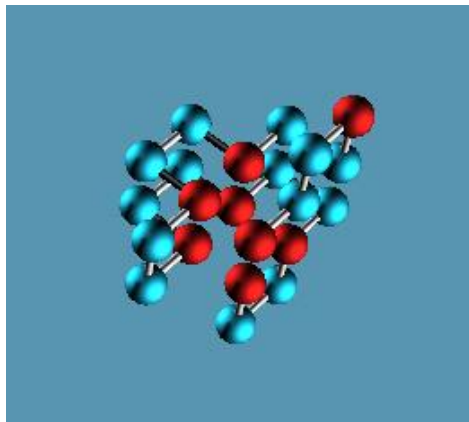
## HP Model

	NONPOLAR, HYDROPHOBIC	R GROUPS	POLAR, UNCHARGED	
Alanine Ala A MW = 89	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_3 \end{array}$		$\text{H} - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array}$	Glycine Gly G MW = 75
Valine Val V MW = 117	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH} \begin{array}{l} \text{CH}_3 \\   \\ \text{CH}_3 \end{array} \end{array}$		$\text{HO} - \text{CH}_2 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array}$	Serine Ser S MW = 105
Leucine Leu L MW = 131	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{CH} \begin{array}{l} \text{CH}_3 \\   \\ \text{CH}_3 \end{array} \end{array}$		$\begin{array}{c} \text{OH} \\   \\ \text{CH}_3 - \text{CH} - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array} \end{array}$	Threonine Thr T MW = 119
Isoleucine Ile I MW = 131	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH} \begin{array}{l} \text{CH}_3 \\   \\ \text{CH}_2 - \text{CH}_3 \end{array} \end{array}$		$\text{HS} - \text{CH}_2 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array}$	Cysteine Cys C MW = 121
Phenylalanine Phe F MW = 131	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{C}_6\text{H}_5 \end{array}$		$\text{HO} - \text{C}_6\text{H}_4 - \text{CH}_2 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array}$	Tyrosine Tyr Y MW = 181
Tryptophan Trp W MW = 204	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{C}_8\text{H}_6\text{N}_2 \end{array}$		$\begin{array}{c} \text{NH}_2 \\   \\ \text{O} = \text{C} - \text{CH}_2 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array} \end{array}$	Asparagine Asn N MW = 132
Methionine Met M MW = 149	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{CH}_2 - \text{S} - \text{CH}_3 \end{array}$		$\begin{array}{c} \text{NH}_2 \\   \\ \text{O} = \text{C} - \text{CH}_2 - \text{CH}_2 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array} \end{array}$	Glutamine Gln Q MW = 146
Proline Pro P MW = 115	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{CH} - \text{CH}_2 \\   \quad   \\ \text{HN} - \text{CH}_2 \end{array}$		<b>POLAR BASIC</b> $\text{NH}_3^+ - \text{CH}_2 - (\text{CH}_2)_3 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array}$	Lysine Lys K MW = 146
Aspartic acid Asp D MW = 133	<b>POLAR ACIDIC</b> $\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{C}(=\text{O})\text{O}^- \end{array}$		$\begin{array}{c} \text{NH}_2 \\   \\ \text{N H}_2^+ = \text{C} - \text{NH} - (\text{CH}_2)_3 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array} \end{array}$	Arginine Arg R MW = 174
Glutamine acid Glu E MW = 147	$\begin{array}{c} ^- \text{OOC} \\   \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{CH}_2 - \text{C}(=\text{O})\text{O}^- \end{array}$		$\begin{array}{c} \text{C} - \text{CH}_2 - \text{CH} - \begin{array}{c} \text{COO}^- \\   \\ \text{N H}_3^+ \end{array} \\   \\ \text{HN}^+ \text{NH} \end{array}$	Histidine His H MW = 155

# HP model



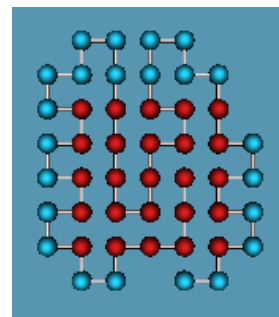
Grid 2D



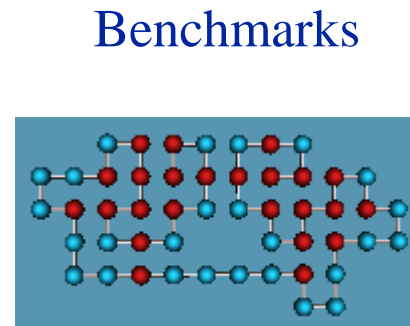
Grid 3D

- Amino acids are classified in:
  - **H (hydrophobic)**: low propensity to be in contact with water, tendency to be buried inside the protein core
  - **P (polar)**: tendency to be in the protein surface in contact with water
- Each protein is represented as a chain:

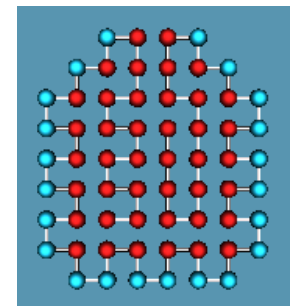
HPHPPHHPHPPHHPHPPH



Opt - 23



Opt - 21



Opt - 42

## Benchmarks

# HP Model. Protein Structure Prediction with Evolutionary Algorithms

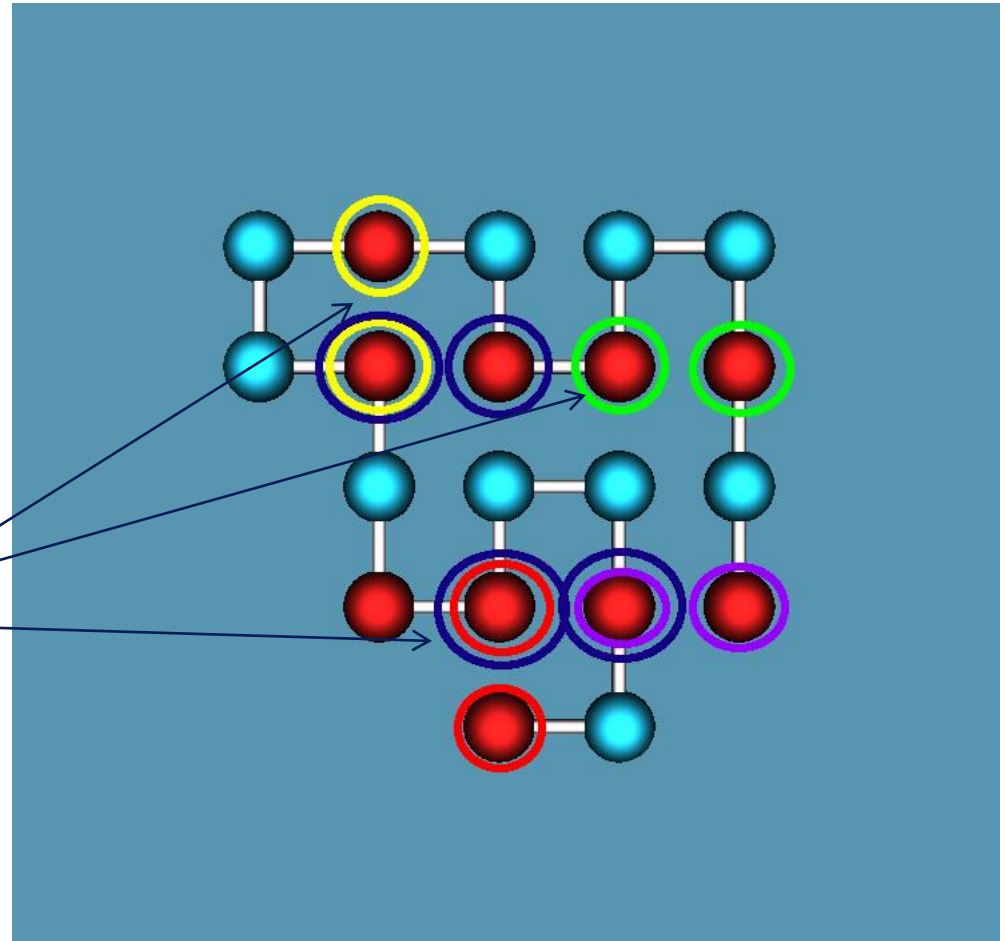
HP energy matrix

$$E = \begin{pmatrix} -1 & 0 \\ 0 & 0 \end{pmatrix}$$

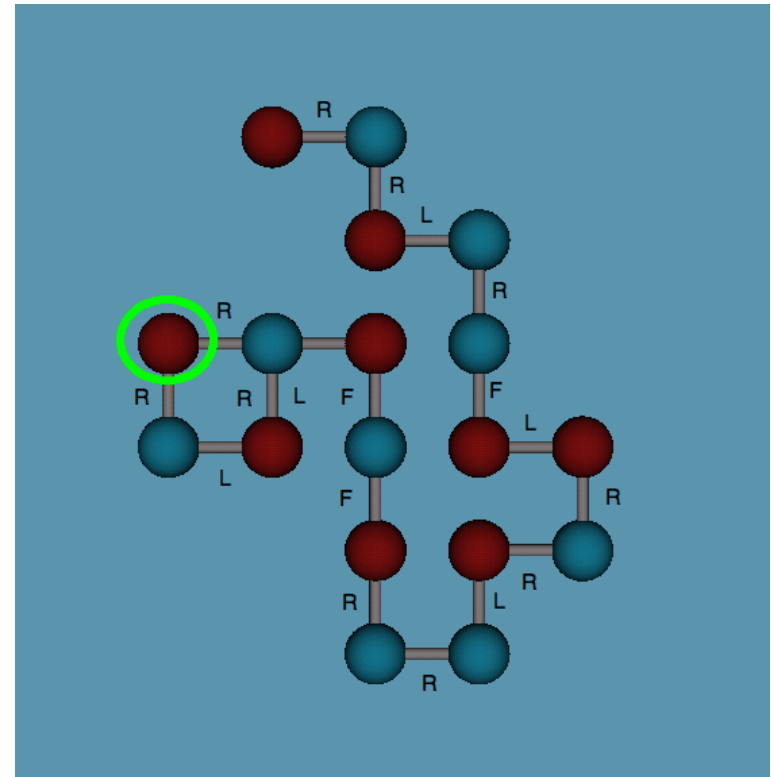
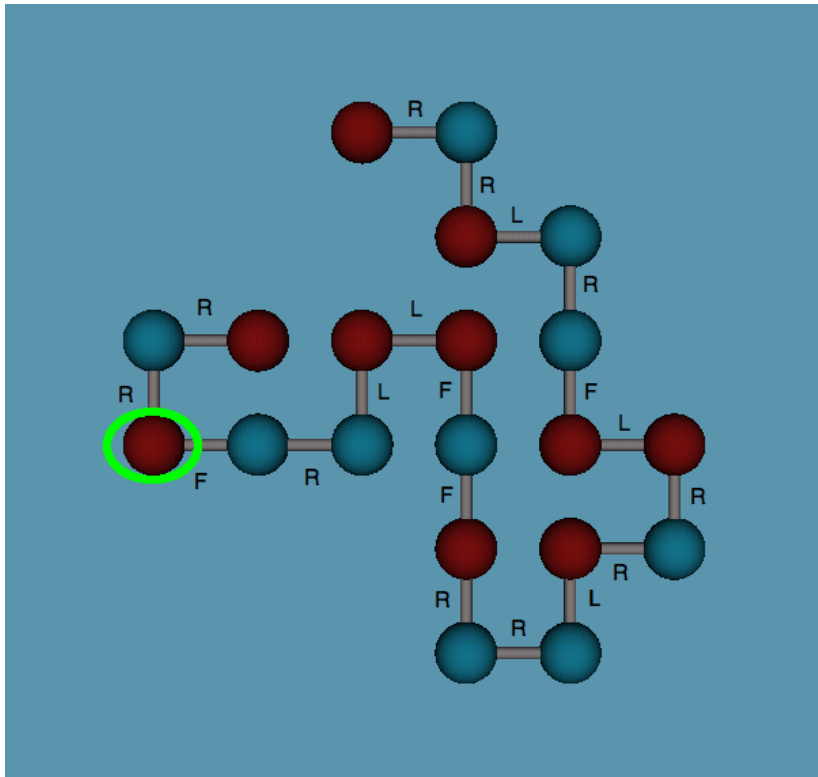
HH contacts

HH contacts

The energy reflects the fact that the hydrophobic amino acids have a propensity to form a hydrophobic inner core.



# Protein Conformation Representation: relative moves



Relative moves: F, L, R (Grid 2D)

# Differential Evolution

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## Algorithm 1 Differential Evolution Algorithm.

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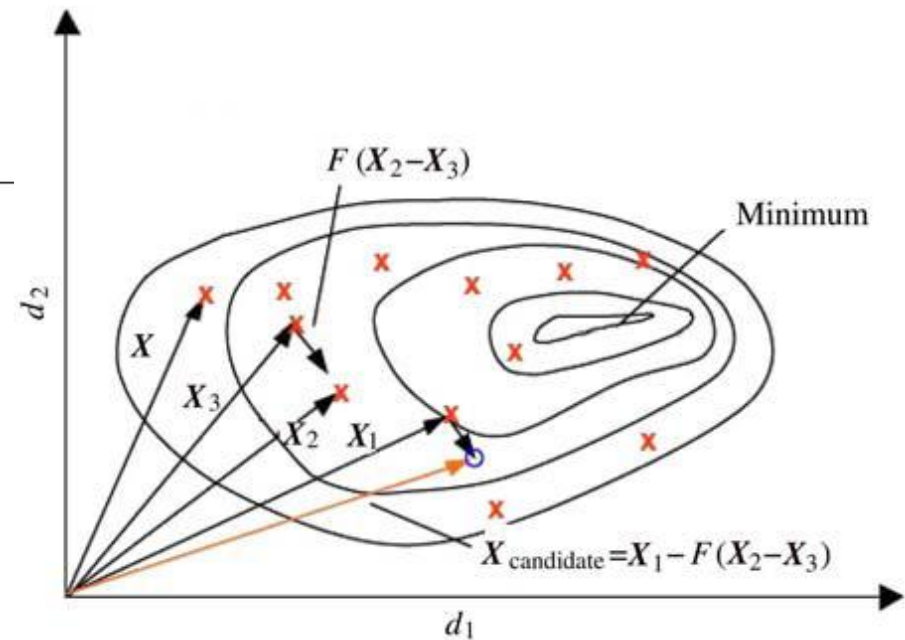
```
1: Initialize the population randomly
2: repeat
3:   for all individual  $x$  in the population do
4:     Let  $x_1, x_2, x_3 \in$  population, randomly obtained  $\{x_1, x_2, x_3, x$  different from each other. $\}$ 
5:     Let  $R \in \{1, \dots, n\}$ , randomly obtained  $\{n$  is the length of the chain. $\}$ 
6:     for  $i = 1$  to  $n$  do
7:       Pick  $r_i \in U(0, 1)$  uniformly from the open range  $(0, 1)$ .
8:       if  $(i = R) \vee (r_i < CR)$  then
9:          $y_i \leftarrow x_{1i} + F(x_{2i} - x_{3i})$ 
10:      else
11:         $y_i = x_i$ 
12:      end if
13:    end for  $\{y = [y_1, y_2 \dots y_n]$  is a new generated candidate individual $\}$ 
14:    if  $f(y) < f(x)$  then
15:      Replace individual  $x$  by  $y$ 
16:    end if
17:  end for
18: until termination criterion is met
19: return  $z \in$  population  $\setminus \forall t \in$  population,  $f(z) \leq f(t)$ 
```

---

In our application:  $F:0.9$

$CR:0.9$

$X_1$  selected with tournament



# Differential Evolution. Encoding

We used relative coordinates. Three movements in 2D: (F)orward, (R)ight and (L)eft

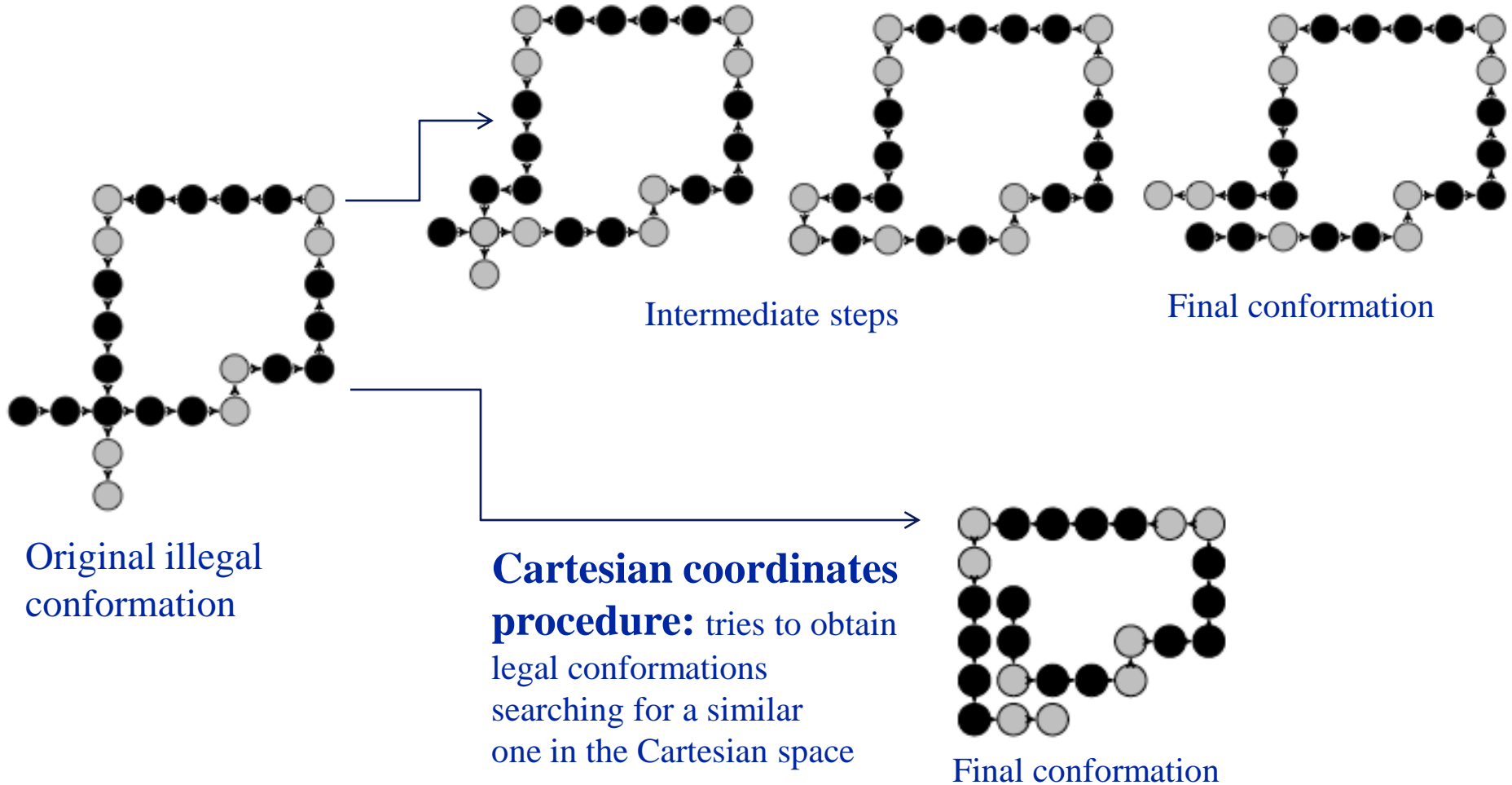
<b>0.24</b>	<b>-0.33</b>	<b>2.44</b>	<b>-1.25</b>	<b>0.18 ...</b>
<b>F</b>	<b>F</b>	<b>R</b>	<b>L</b>	<b>F</b>

movement **L** if  $X_{ij} \in [\alpha, \beta)$   
**F** if  $X_{ij} \in [\beta, \delta)$   
**R** if  $X_{ij} \in [\delta, \gamma]$

$\alpha < \beta < \delta < \gamma$  arbitrary constants in  $\mathbb{R}$  ( $\alpha=-3, \beta=-1, \delta=1, \gamma=3$ )

# Repair process

**Absolute moves procedure:** tries to maintain the relative conformation of the rest of the chain

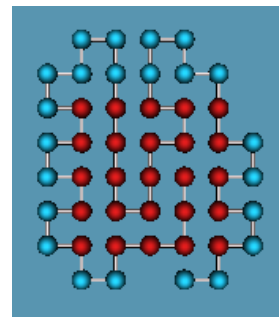


# Some results

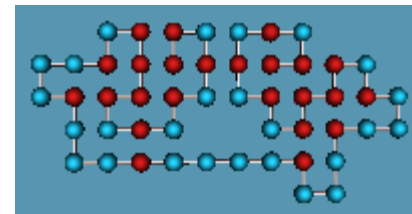
Comparison of results with the benchmark sequences

<i>Seq.</i>	$E_{min}$	U&M GA [23]	hybrid DE	L&B DE [13]	ACO [18]
S1	-9	<b>-9</b> (30492)	<b>-9,-9</b> (3584, 6362)	<b>-9,-9</b>	<b>-9</b>
S2	-9	<b>-9</b> (30491)	<b>-9,-9</b> (5806, 9292)	<b>-9,-9</b>	<b>-9</b>
S3	-8	<b>-8</b> (20400)	<b>-8,-8</b> (7061, 18828)	<b>-8,-8</b>	<b>-8</b>
S4	-14	<b>-14</b> (301339)	<b>-14,-14</b> (45793, 92579)	<b>-14,-13.96</b>	<b>-14</b>
S5	-23	<b>-22</b> (126547)	<b>-23,-23</b> (245943, 532787)	<b>-23,-23</b>	<b>-23</b>
S6	-21	<b>-21</b> (592887)	<b>-21,-21</b> (365222, 691989)	<b>-21,-21</b>	<b>-21</b>
S7	-36	<b>-34</b> (208781)	<b>-35,-33.57</b>	<b>-35,-34.79</b>	<b>-36</b>
S8	-42	<b>-37</b> (187393)	<b>-42,-42</b> (176313, 340917)	<b>-42,-41.87</b>	<b>-42</b>

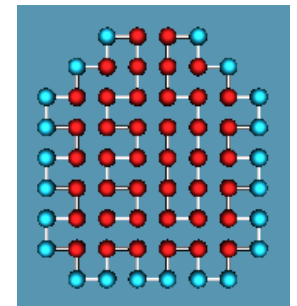
Optimal configurations for three of the sequences: S5, S6 and S8



S5 - 23



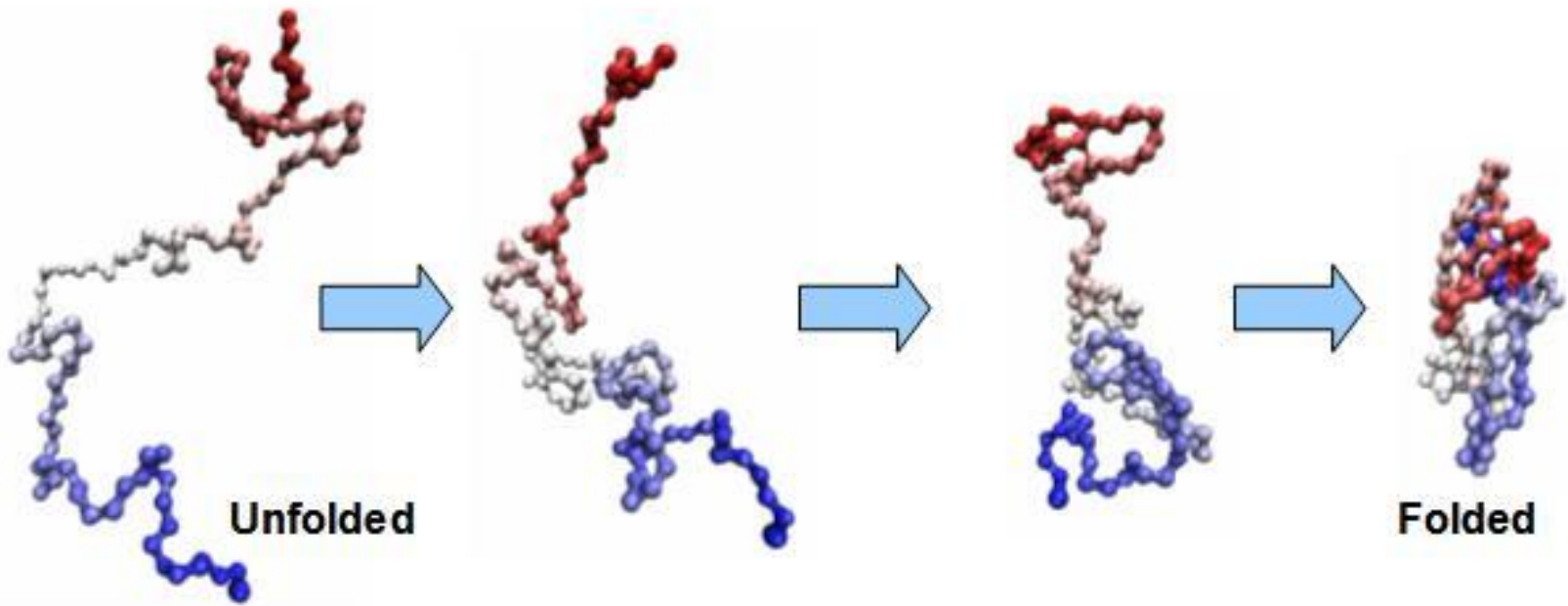
S6 - 21



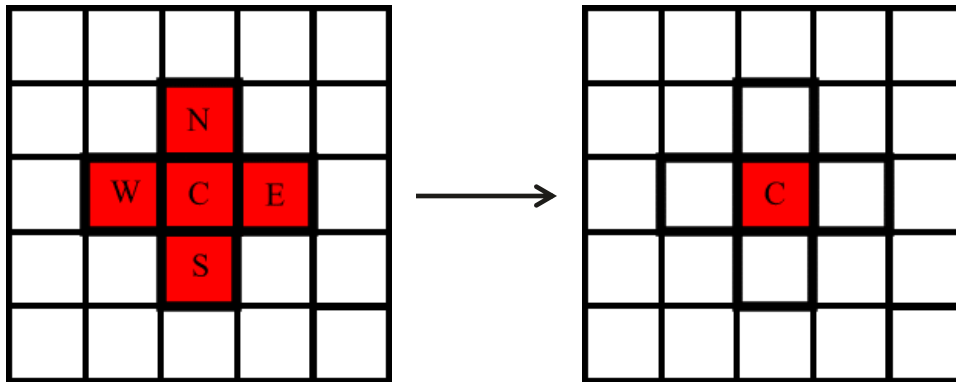
S8 - 42



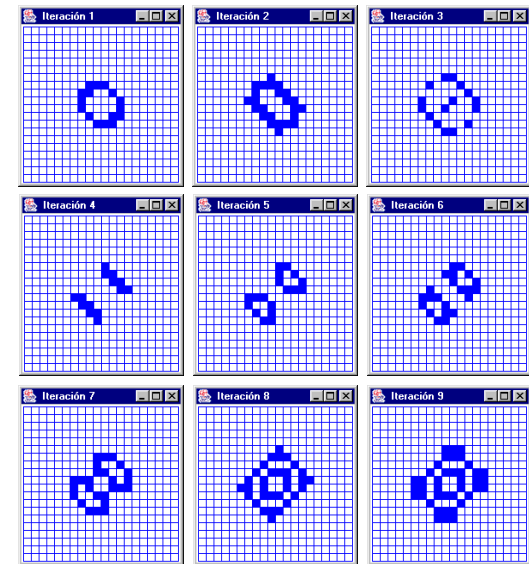
# Protein folding



# Cellular automata



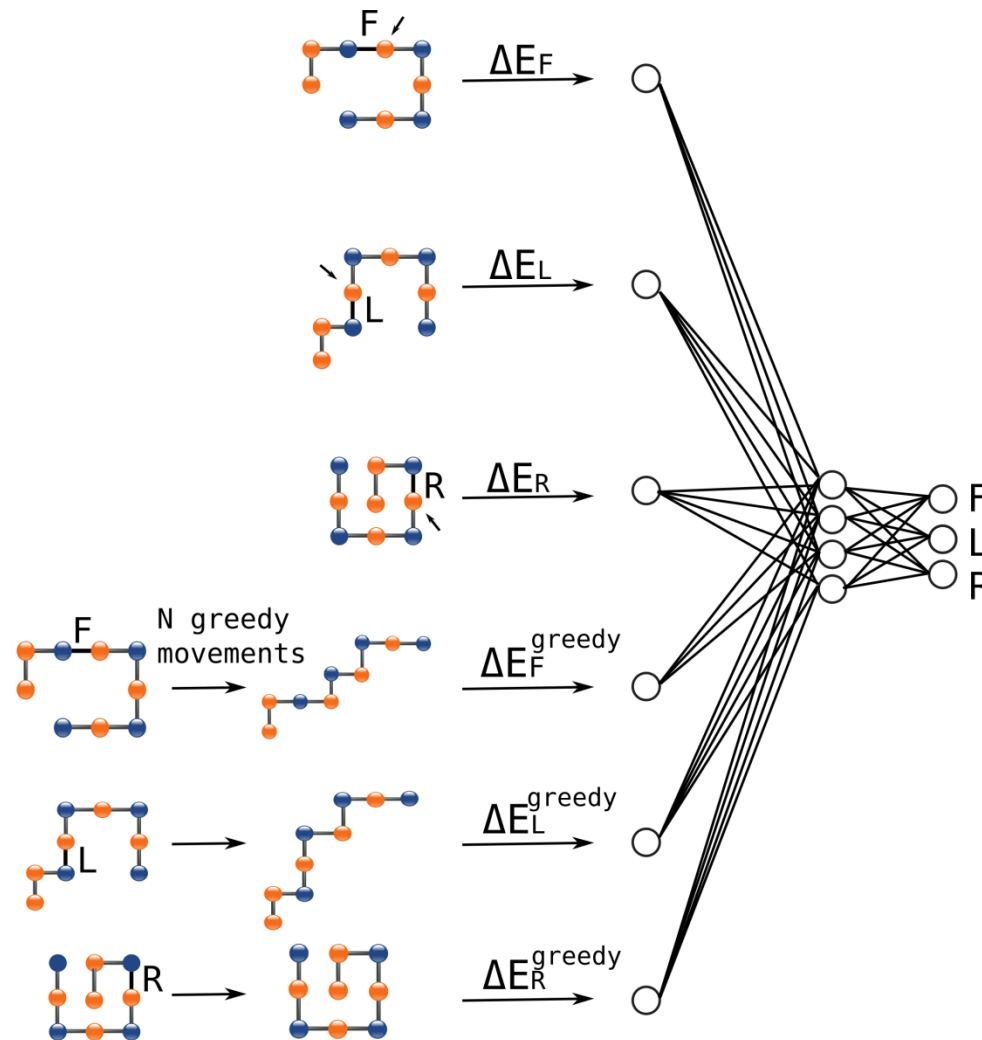
von Neumann neighborhood in a cellular automaton in 2D



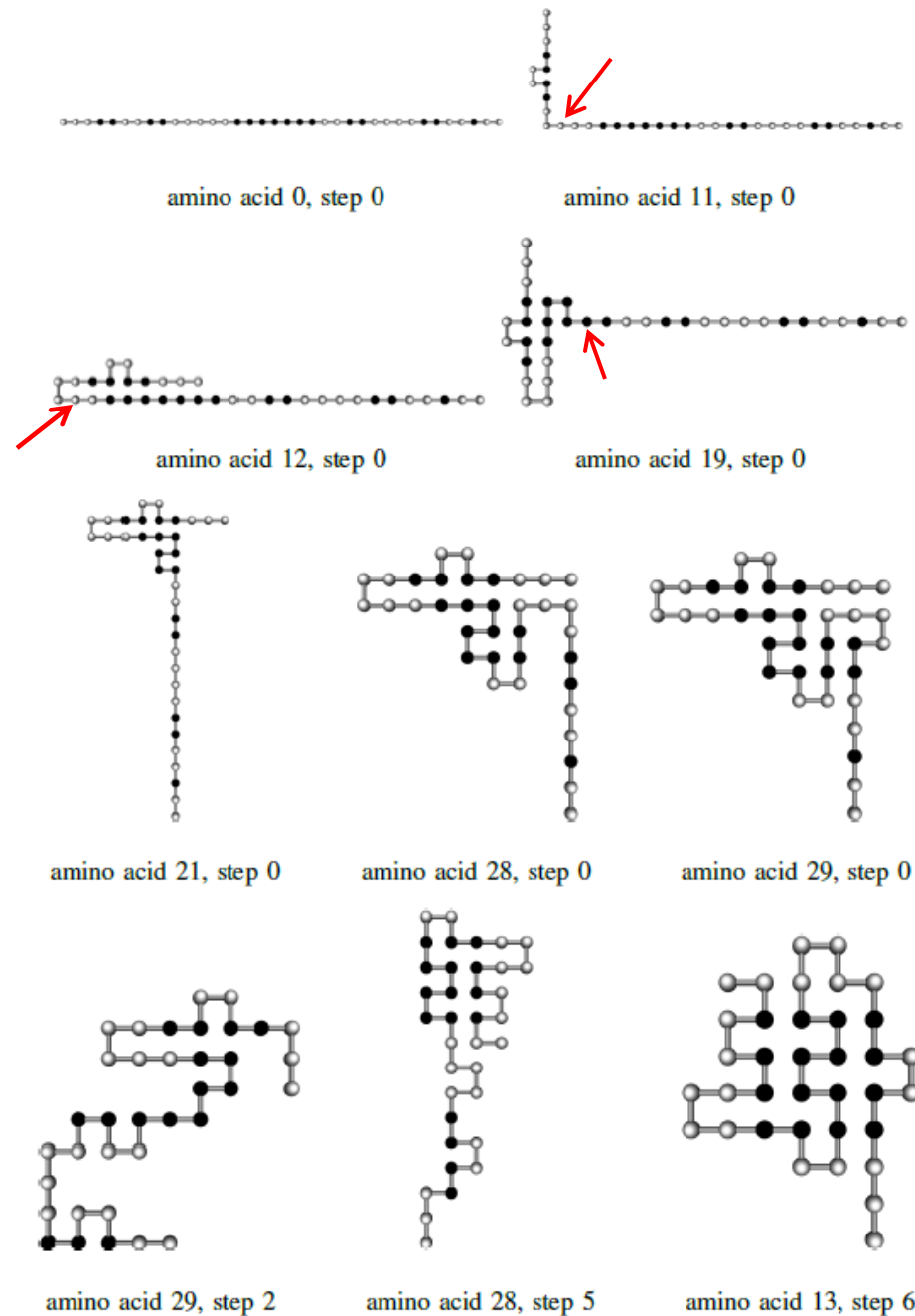
Conway's "Game of Life"



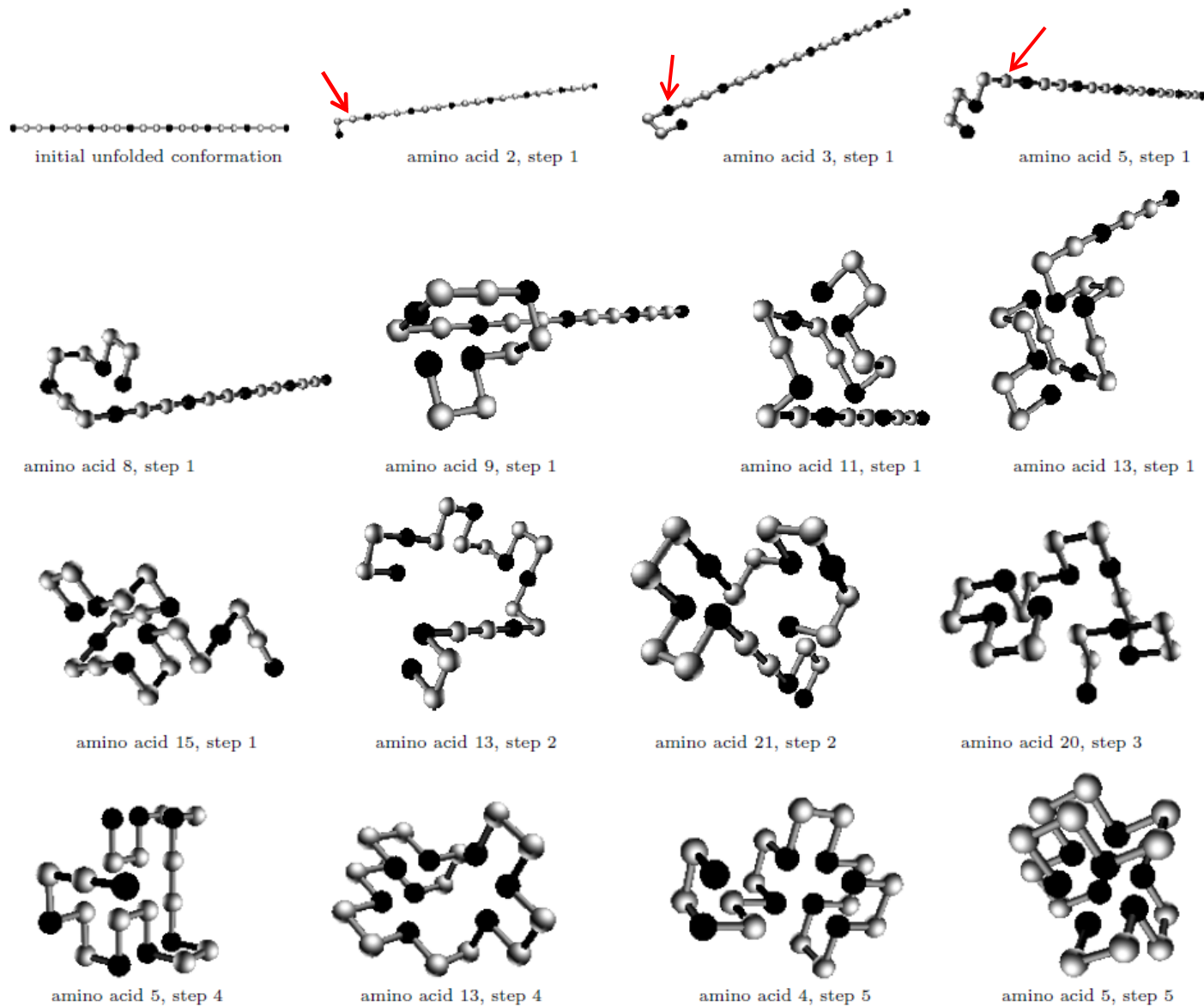
# Protein folding with “Neural Cellular Automata”



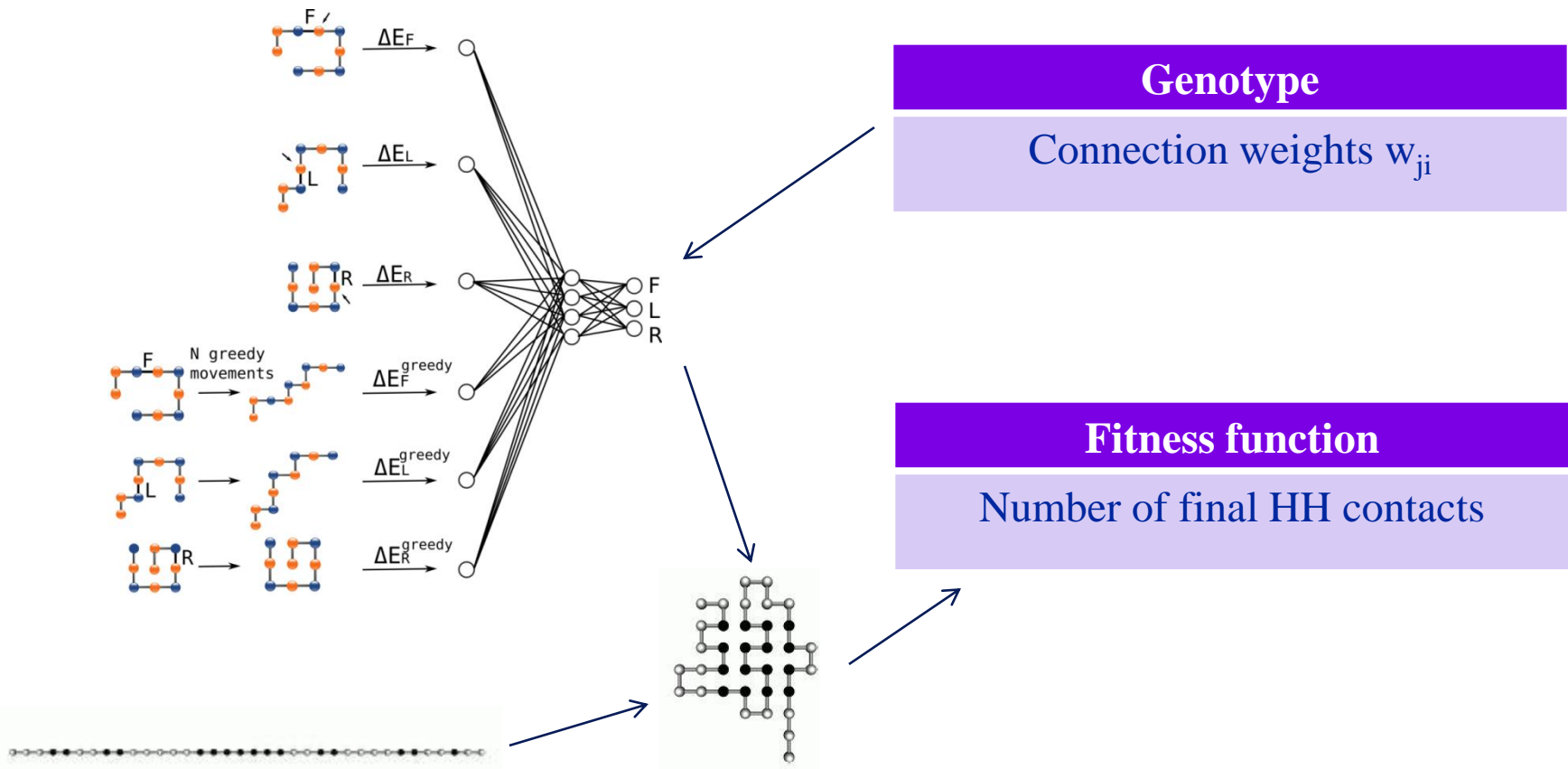
# Protein folding. Example in 2D with the HP model



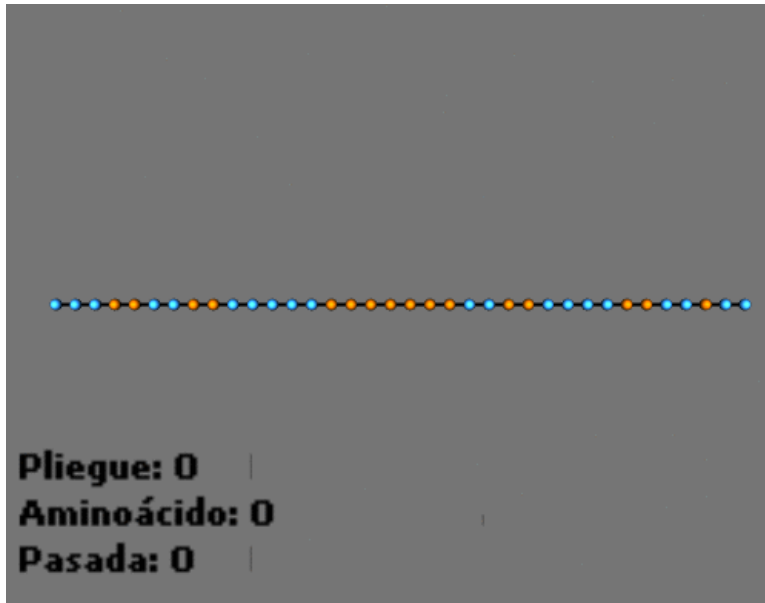
# Protein folding. Example in 3D with the HP model



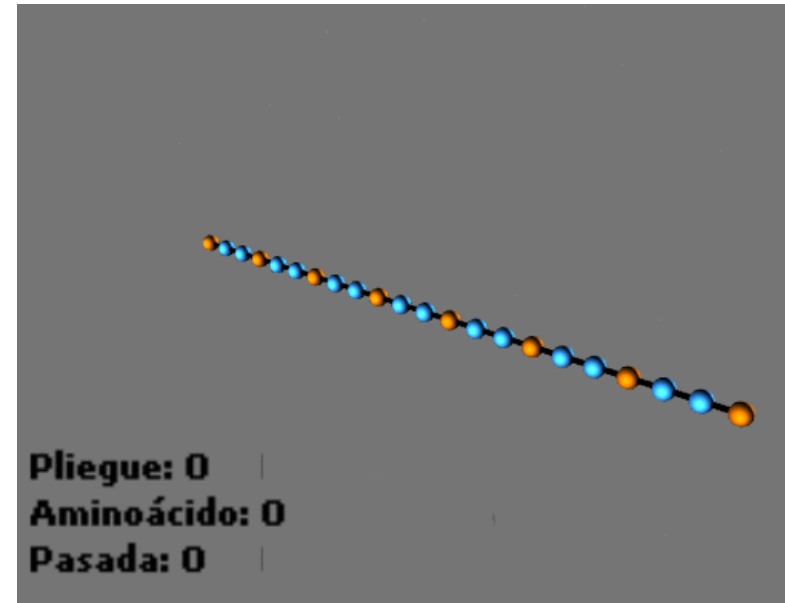
# Evolving “Neural Cellular Automata” with Differential Evolution



# Protein folding process modeling with the HP model

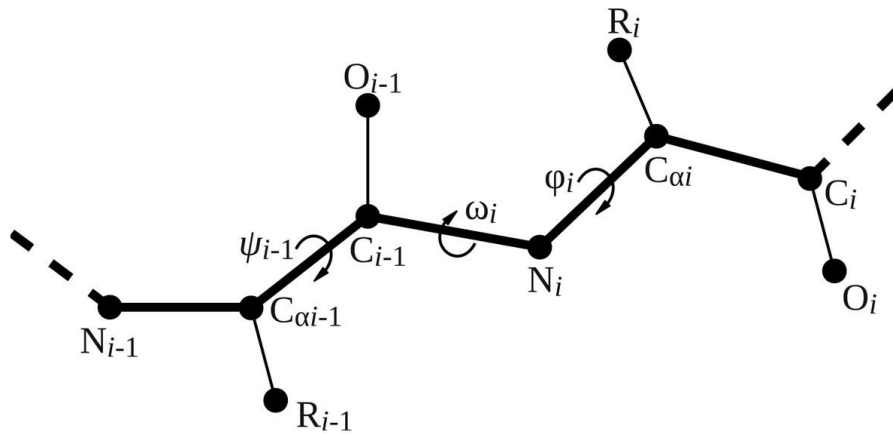


2D sequence – Optimum: 14 HH contacts

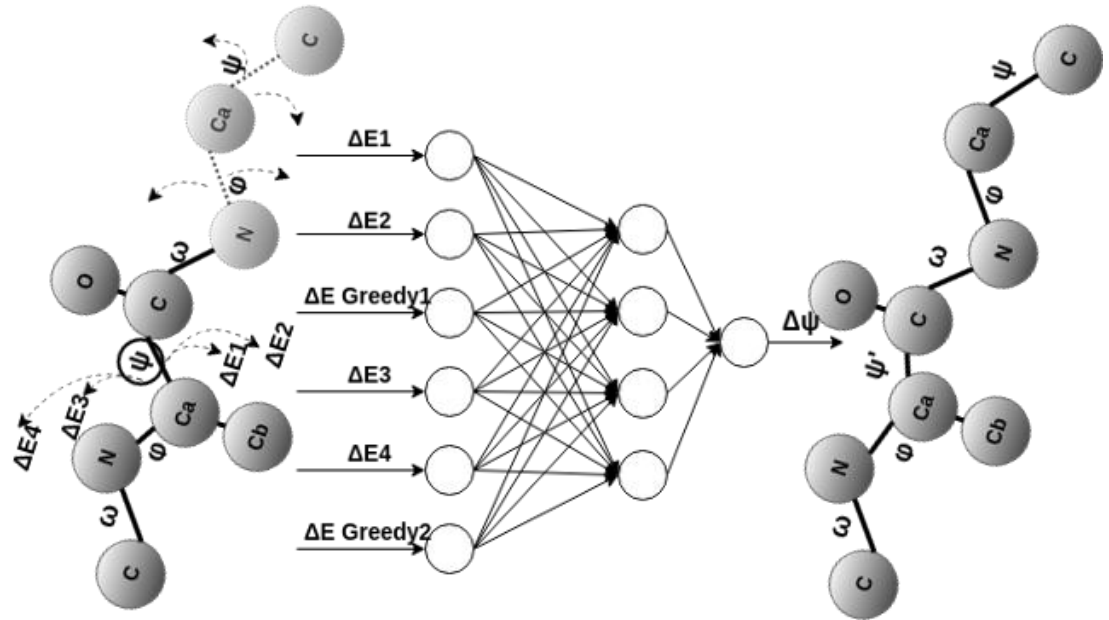
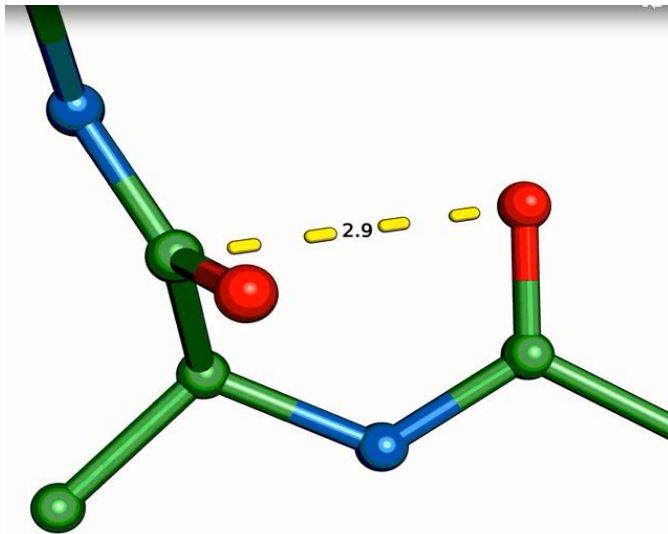


3D sequence – Optimum: 12 HH contacts

# Protein folding process modeling with the off-lattice model of the Rosetta system



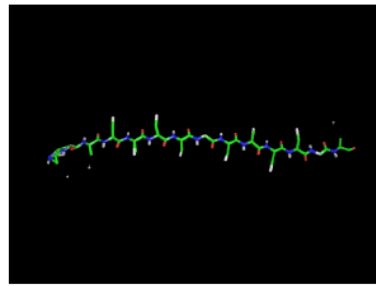
Protein conformation representation with the dihedral angles



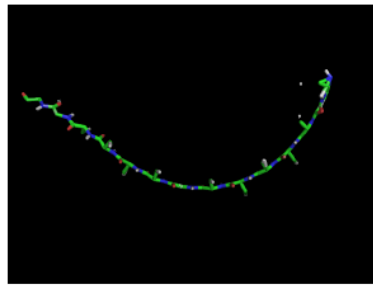
Neural cellular automaton



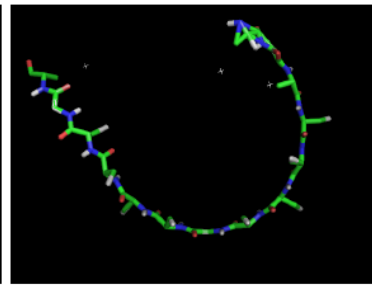
# Protein folding process modeling with the off-lattice model of the Rosetta system



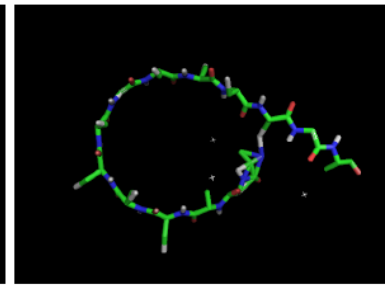
initial unfolded conformation



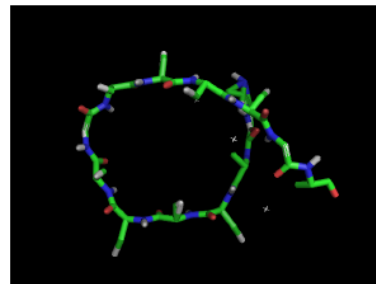
end of step 1



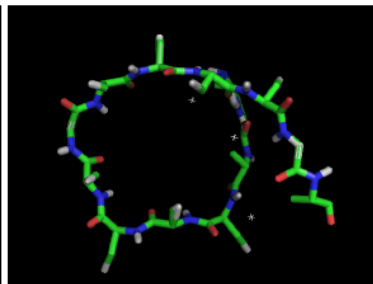
end of step 2



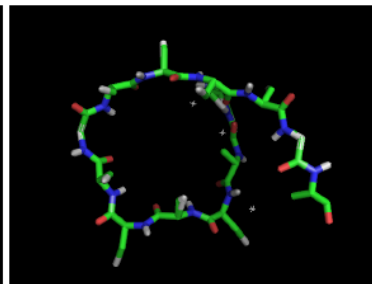
end of step 3



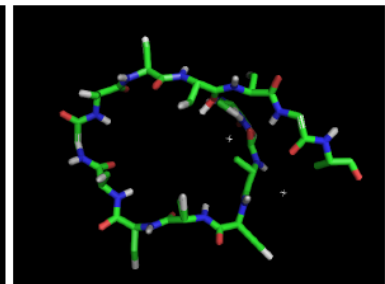
end of step 6



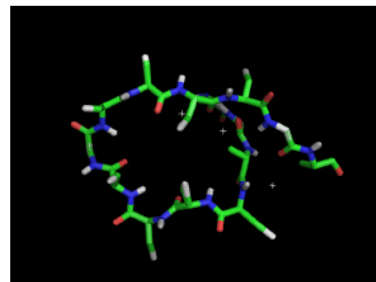
end of step 8



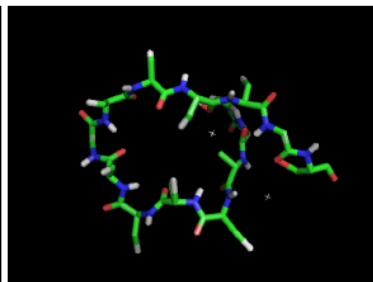
end of step 10



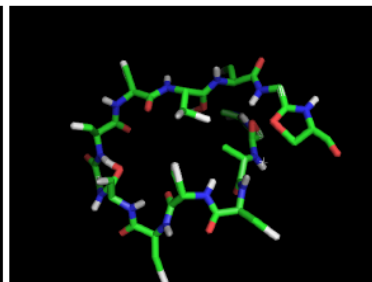
end of step 12



end of step 14



end of step 16



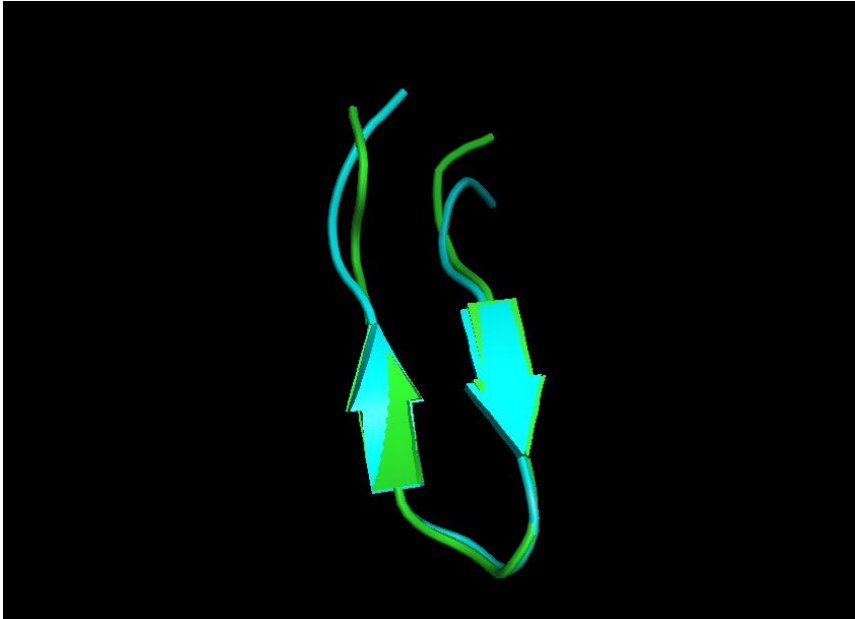
end of step 19



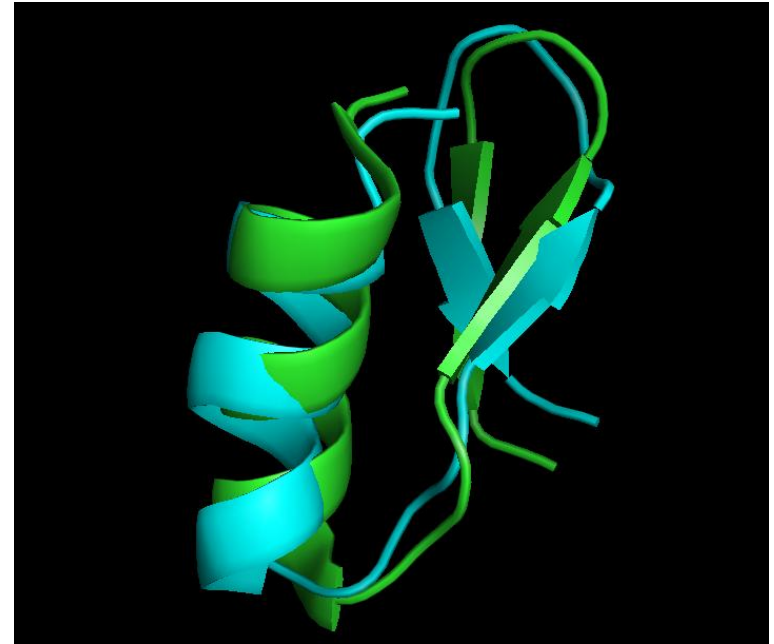
native and final folded structure

Snapshots of the folding at the end of different temporal steps with protein sequence 1j4m. The last snapshot shows the native structure (green) and the final folded structure at the last step (blue).

# Protein folding process modeling with the off-lattice model of the Rosetta system



**Final folded conformation for protein 1j4m (14 amino acids) (blue) and the corresponding native structure (green) when the energy term associated with secondary structure elements is taken into account and when the folding process begins with a partially folded conformation.**



**Final folded conformation for protein 1d5q (27 amino acids) (blue) and the corresponding native structure (green) when the folding process begins with a partially folded conformation.**

# Tumor growth modeling with cellular automata

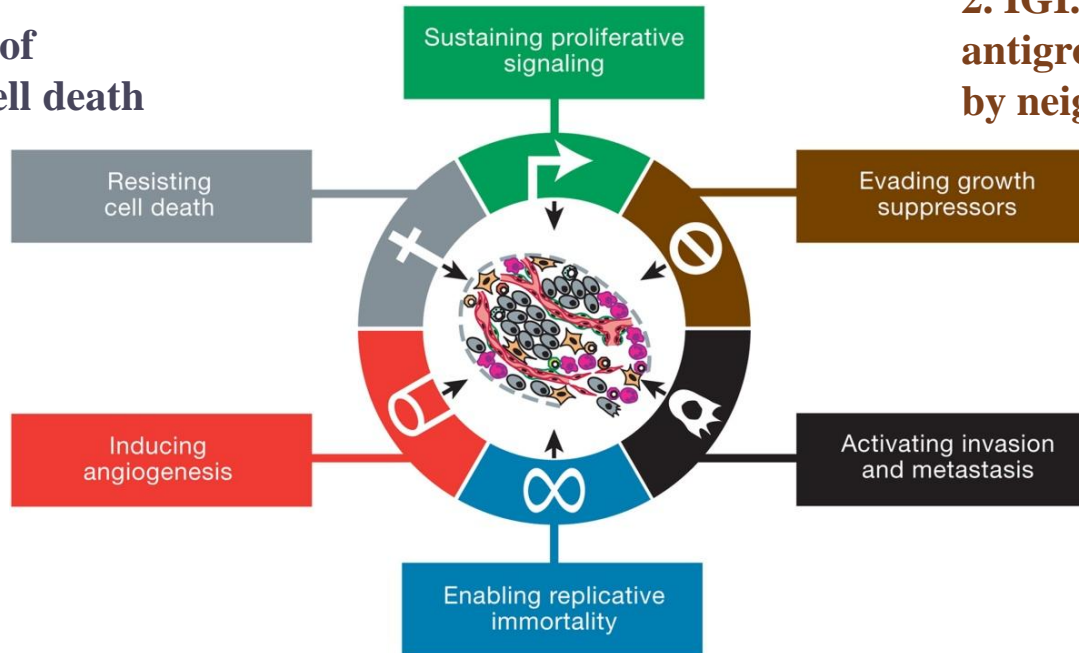
# Cancer hallmarks

**1. SG. Growth even in the absence of normal “go” signals.**

**2. IGI. Growth despite antigrowth signals issued by neighboring cells.**

**3. EA. Evasion of programmed cell death (apoptosis)**

**6. AG. Ability to stimulate blood vessel construction (angiogenesis)**



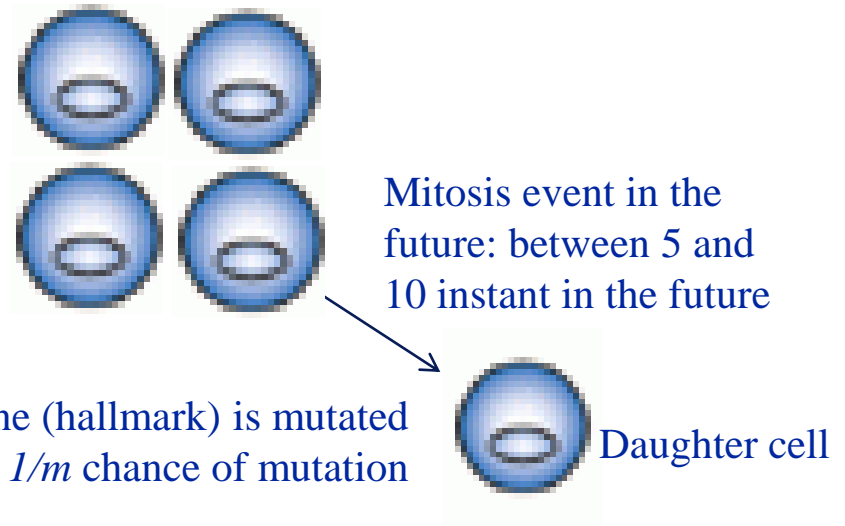
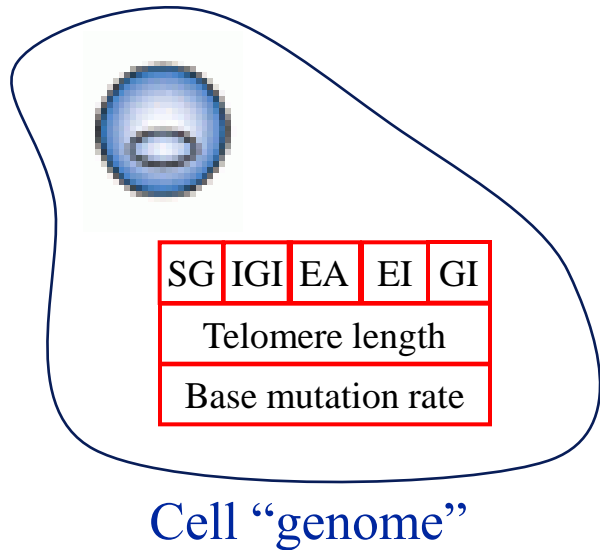
**4. EI. Effective immortality.**

**5. GI. Genetic Instability.**

**7. MT. Power to invade other tissues and spread to other organs.**

Hanahan, D. and Weinberg, R.A. (2000) The hallmarks of cancer, *Cell*, 100, 57-70.

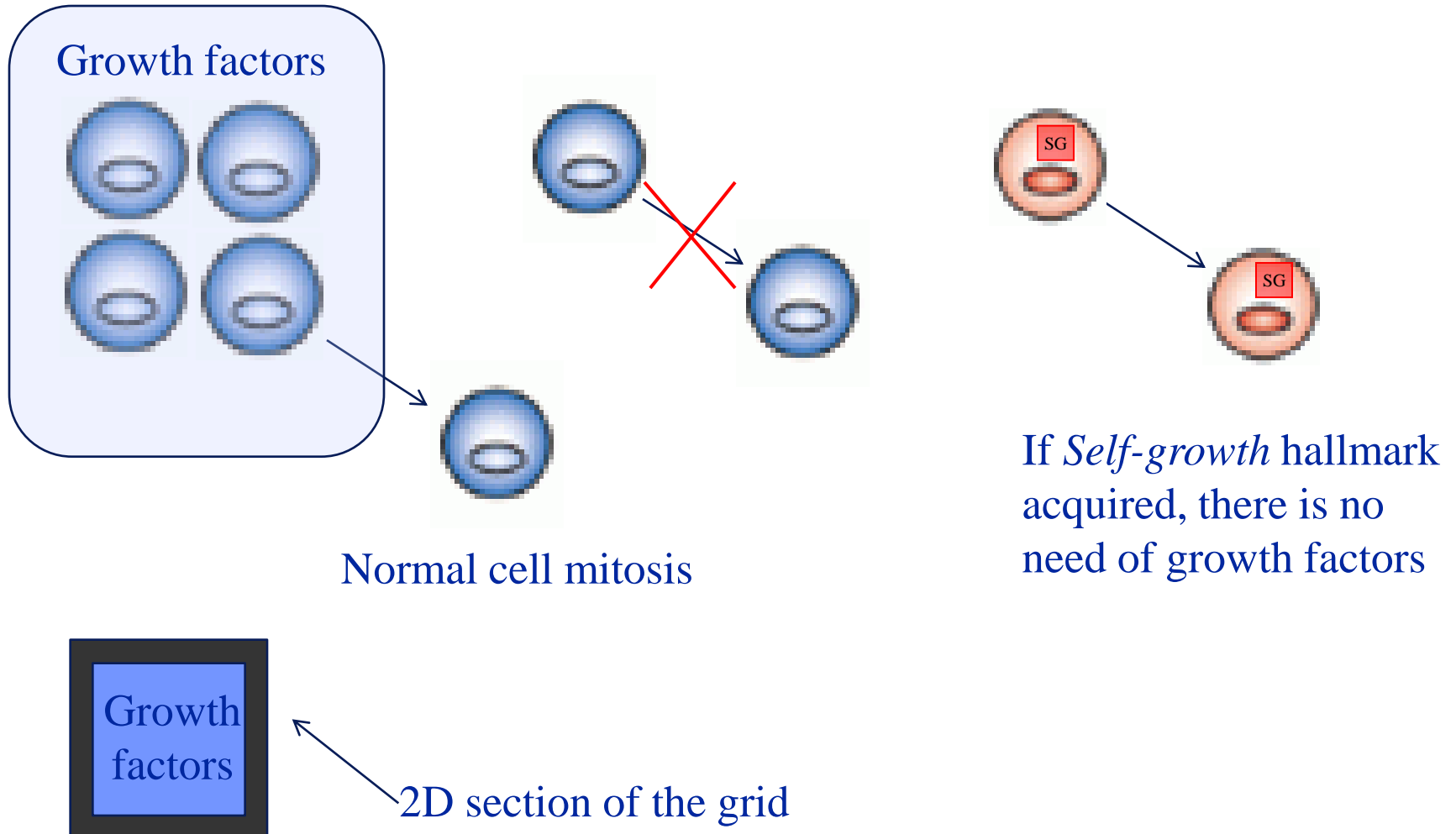
# AC Cell growth simulation



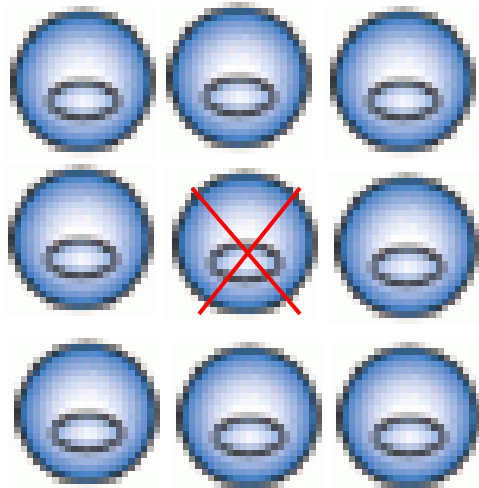
Normal cell mitosis

1 iteration = 2.6 hours  
5000 iterations = 75 weeks

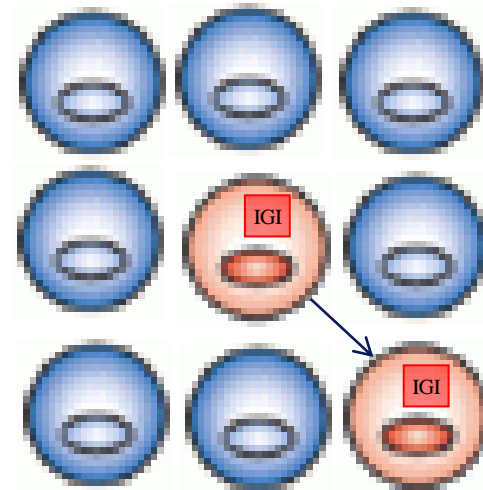
# AC Cell growth simulation. Self-growth hallmark (SG)



# AC Cell growth simulation. Ignore growth inhibit hallmark (IGI)

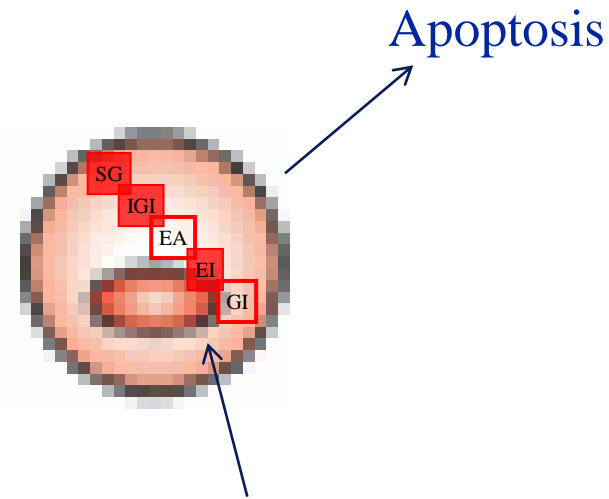


No empty space in the neighborhood

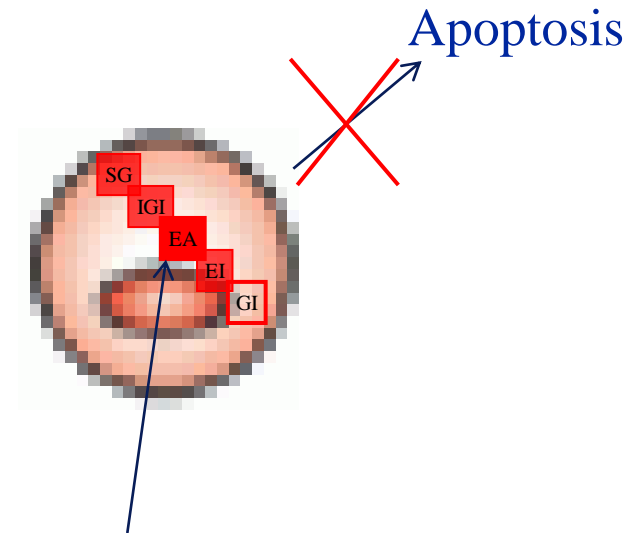


If *Ignore growth inhibit* hallmark acquired, a neighbor is killed (with probability  $1/g$ ) to make room for mitosis

# AC Cell growth simulation. Evade apoptosis hallmark (EA)



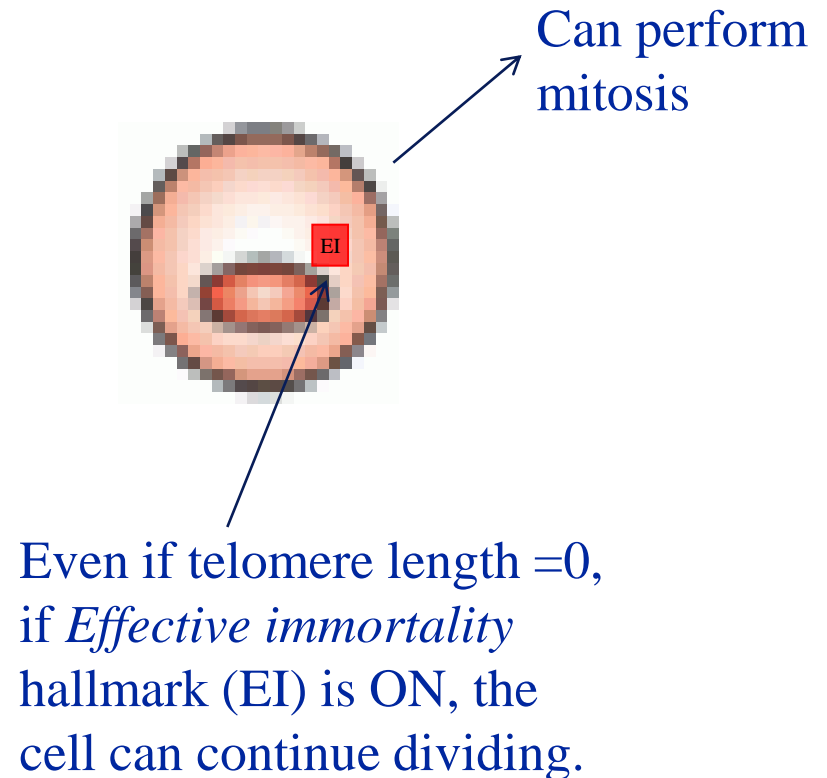
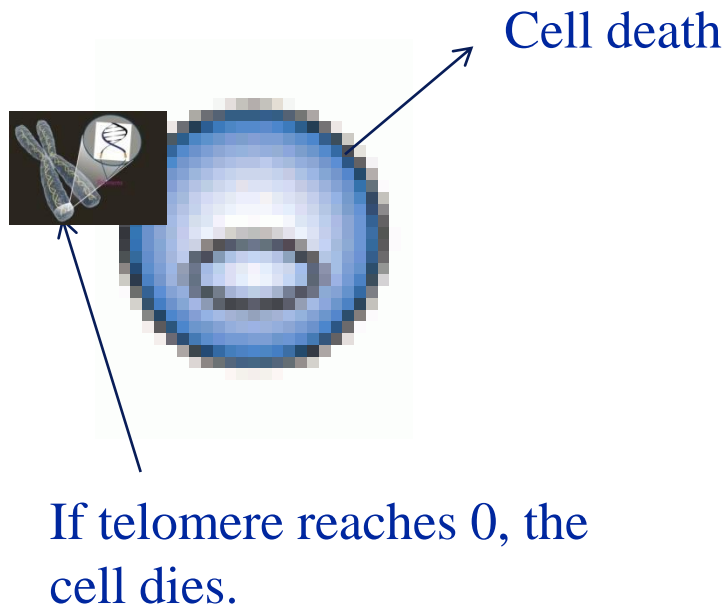
A cell with  $n$  hallmarks mutated has an extra  $n/e$  likelihood of dying each cell cycle.



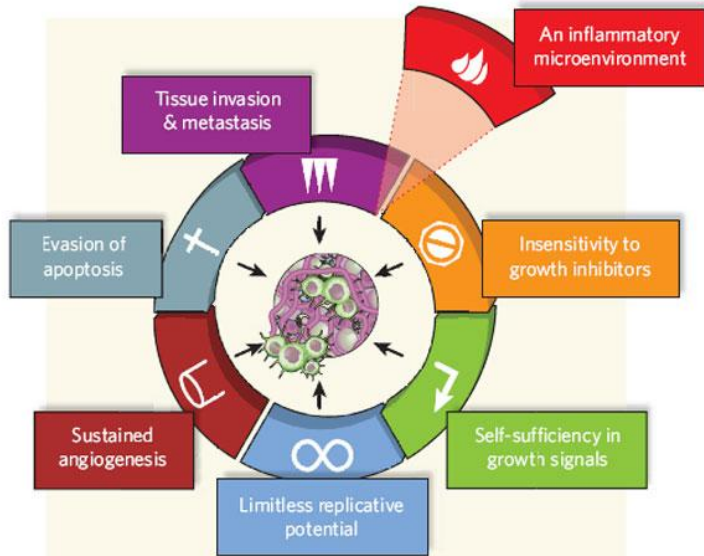
*If Evade apoptosis hallmark (EA) is ON, there is not apoptosis*



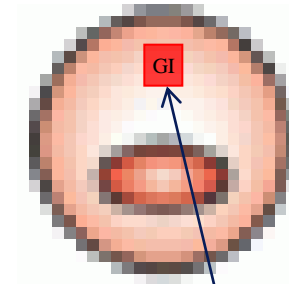
# AC Cell growth simulation. Effective immortality hallmark (EI)



# AC Cell growth simulation. Genetic instability (GI)



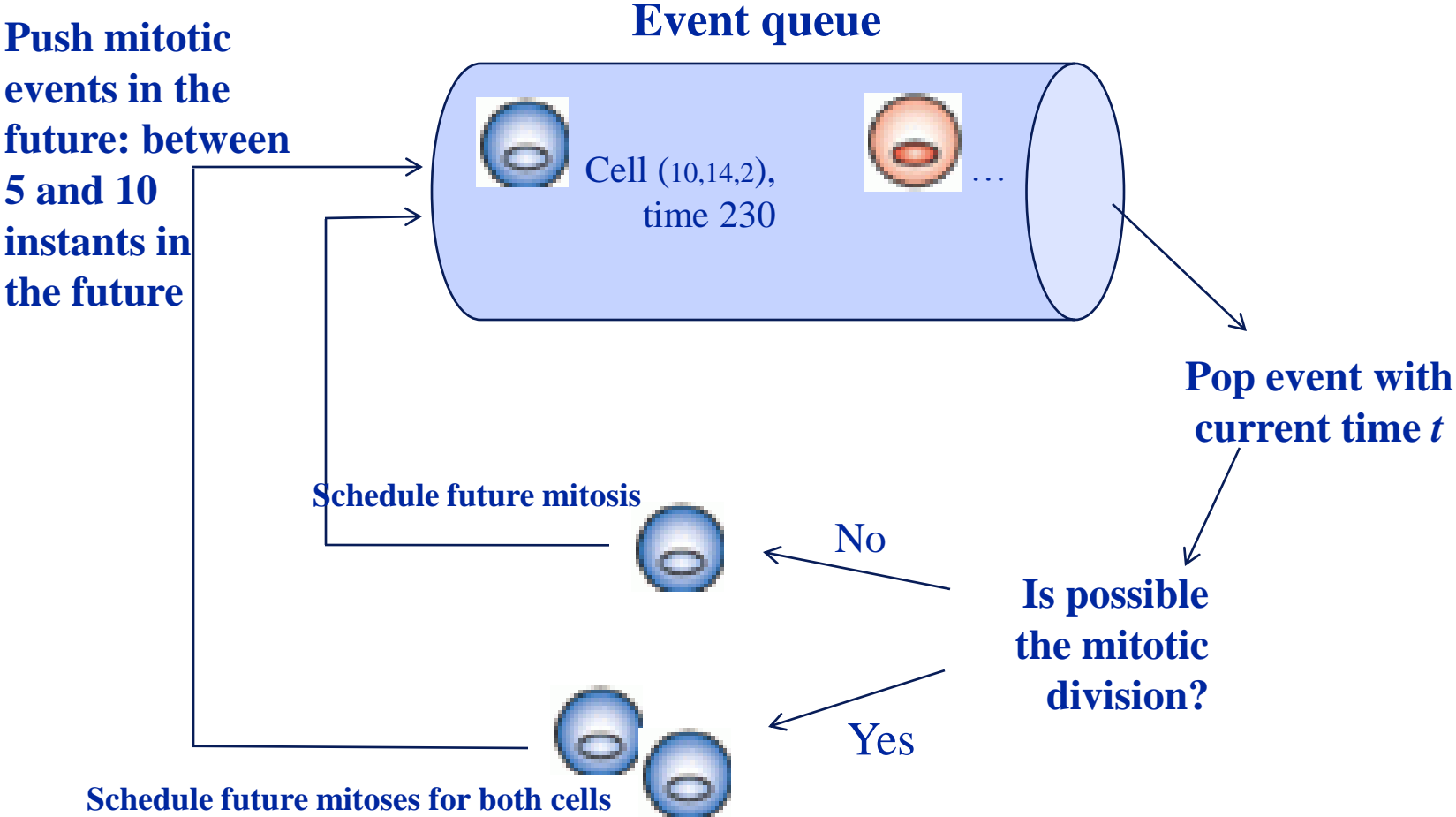
Hanahan and Weinberg (2011), “Hallmarks of Cancer: The Next Generation”, *Cell*.



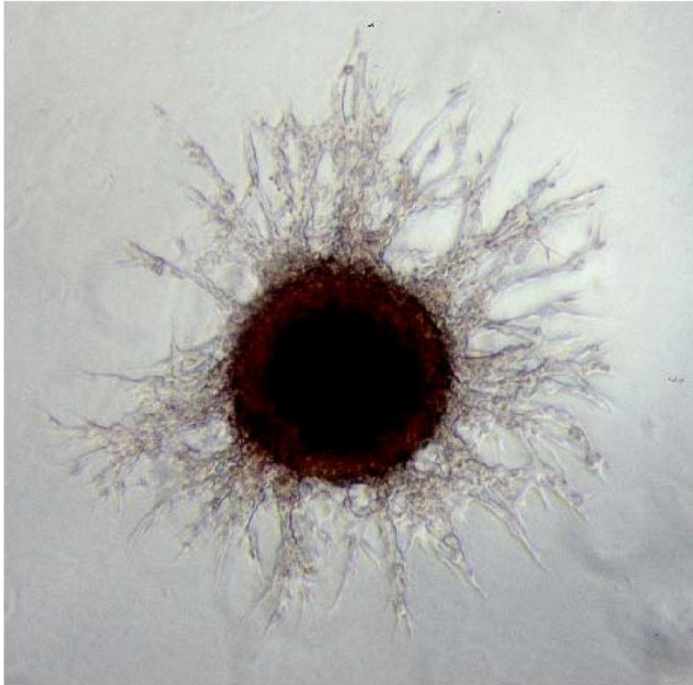
If *Genetic instability* factor (GI) is ON

Increment the cell base mutation rate by a factor ( $i$ )

# Event model

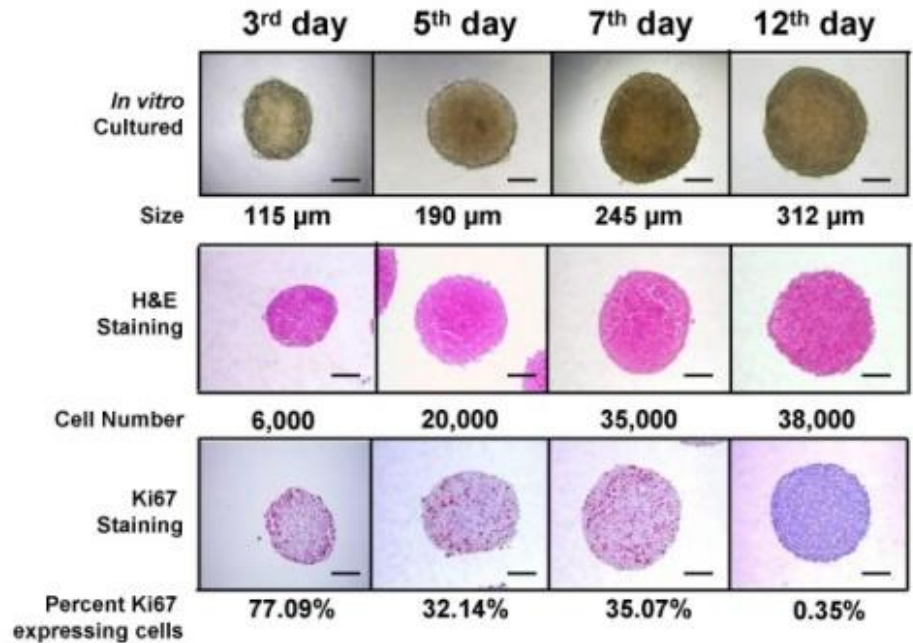


# Simulating cancer cell cultures: Multicellular spheroids



Microscopy image of a multicellular tumor spheroid, exhibiting an extensive branching system that rapidly expands into the surrounding extracellular matrix gel. These branches consist of multiple invasive cells.

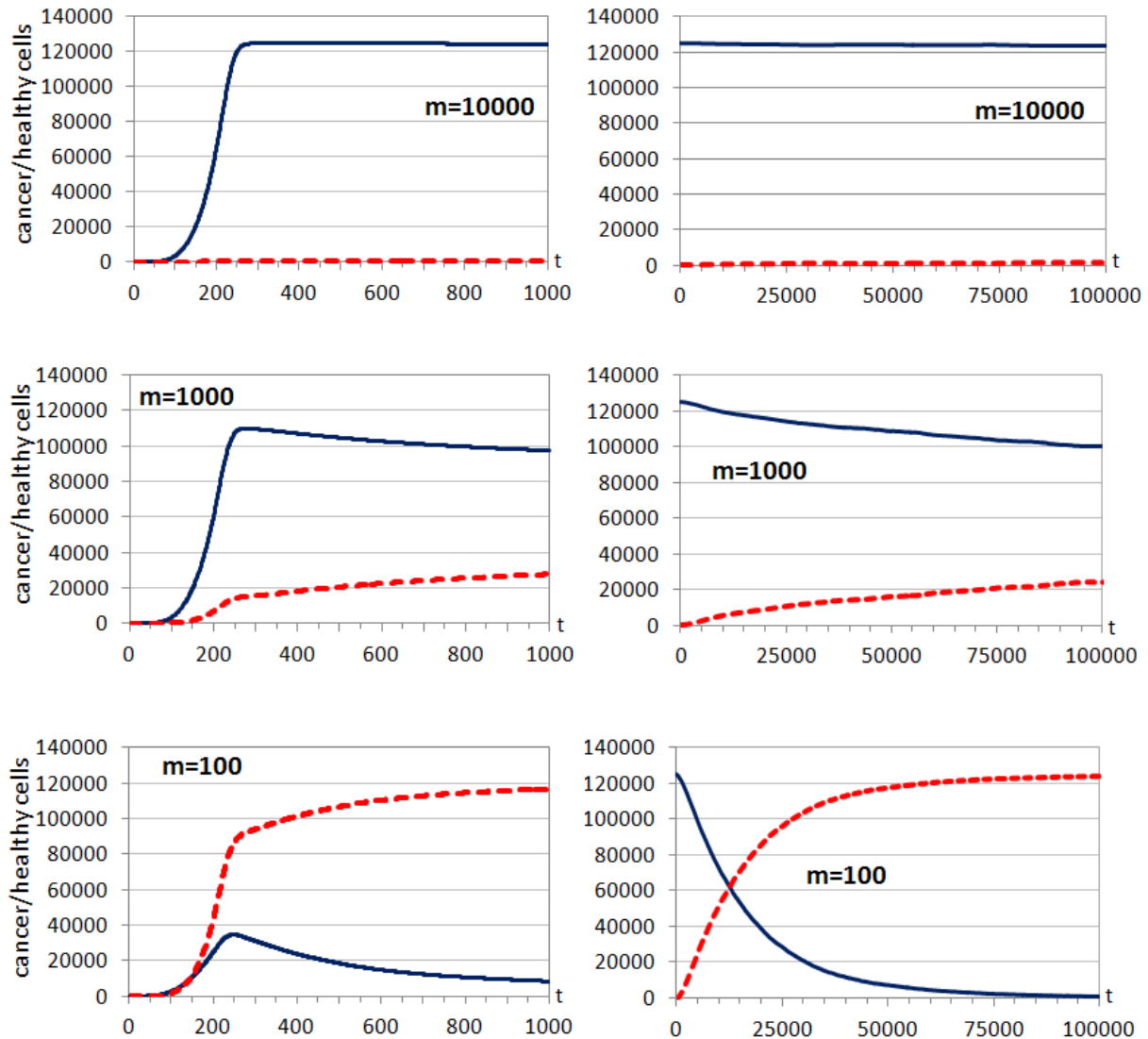
Guiot *et al.* *Theoretical Biology and Medical Modelling* 2007 4:4 doi:10.1186/1742-4682-4-4



Growth stages of 3D-cultured CT26 colon cancer spheroid. Five hundred suspension cancer cells were dispensed into each well of a 48-well culture tray. Trays were then inverted and incubated during 12 days. Spheroids were collected on days 3, 5, 7 and 12 and processed for cell counting, spheroid diameter determination and immunohistochemical detection of Ki67-expressing cells. Scale bar: 100  $\mu\text{m}$ .  
Valcárcel M, et al.- *J Transl Med* (2008)

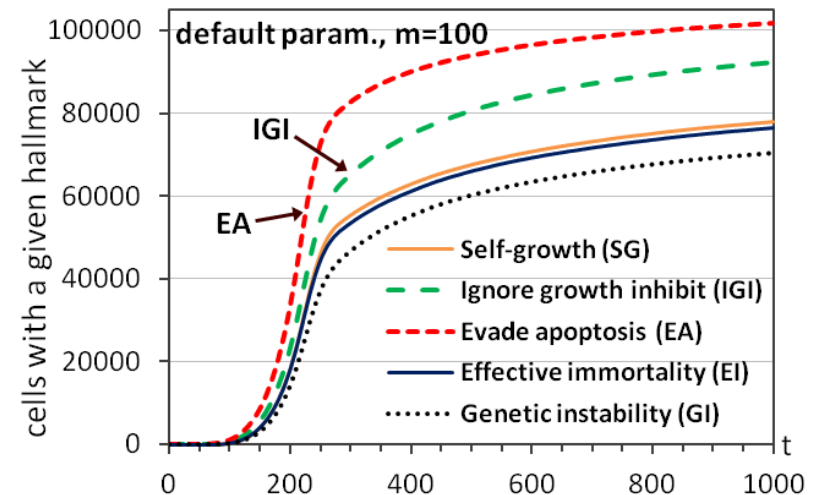
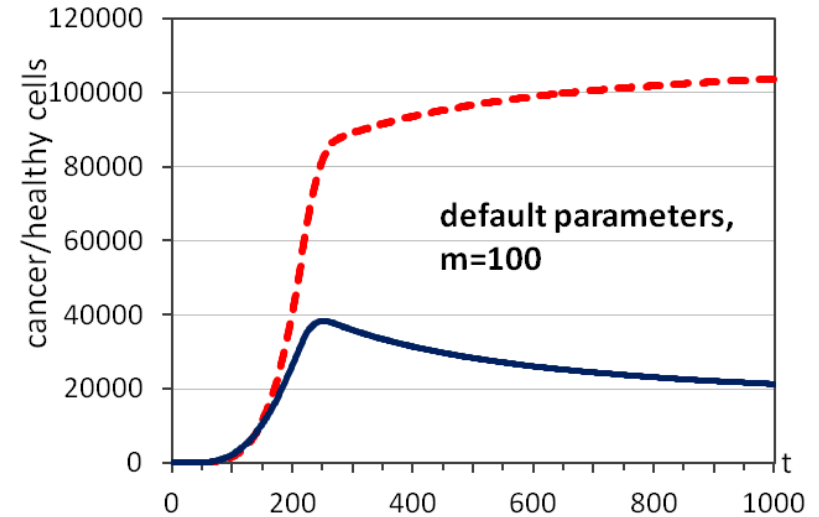
# Some simulations and results

# Dependence on hallmark parameters



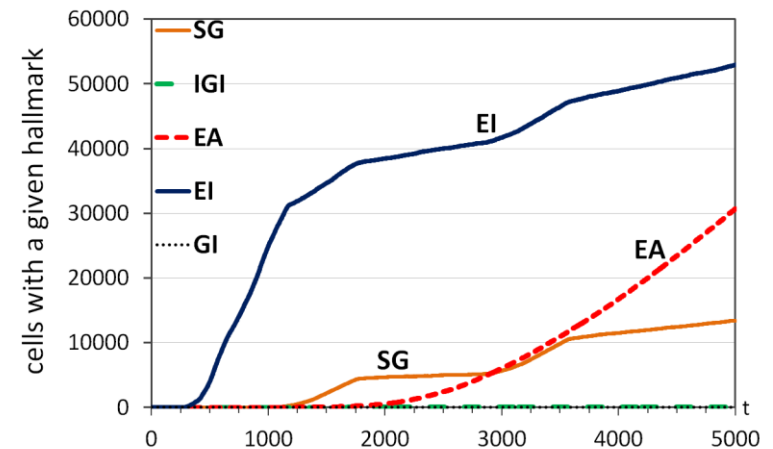
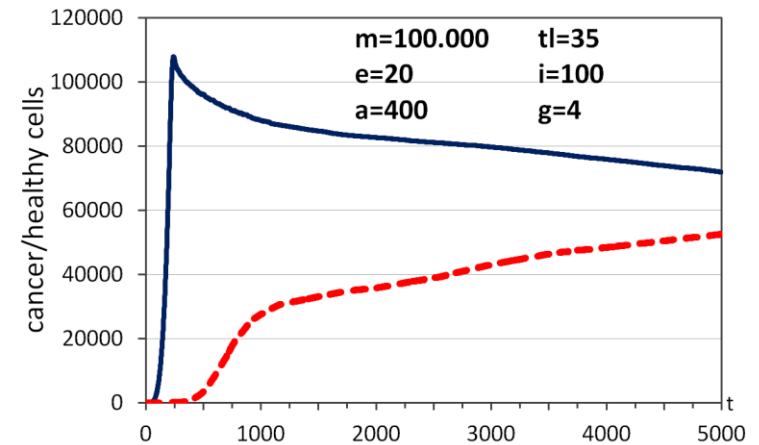
Evolution through time iterations of the number of healthy cells (continuous lines) and cancer cells (dashed lines) for different base mutation rates ( $1/m$ )

# Simulation run with default parameters and $m=100$



Grid size=125000, mutation rate=0.01

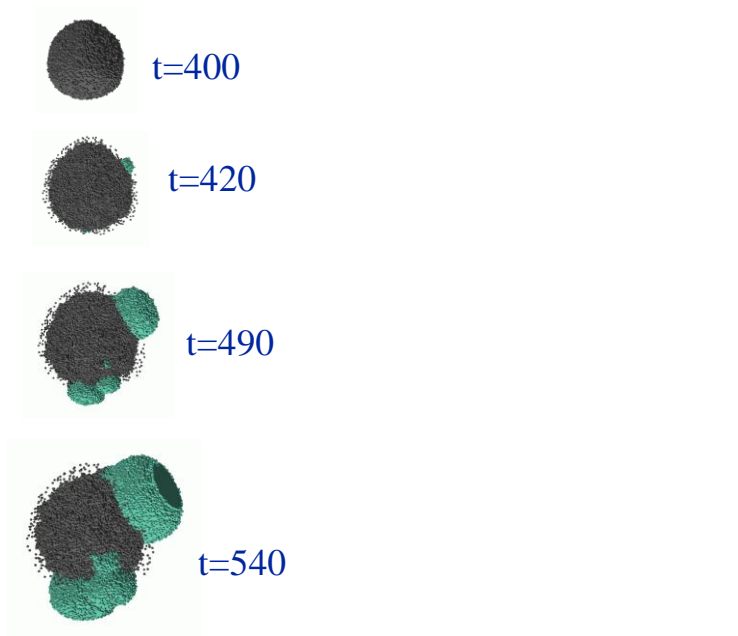
# Simulation run with parameters that facilitates cancer appearance



Grid size=125000, mutation rate= $10^{-5}$



# Simulation run with parameters that facilitates cancer appearance

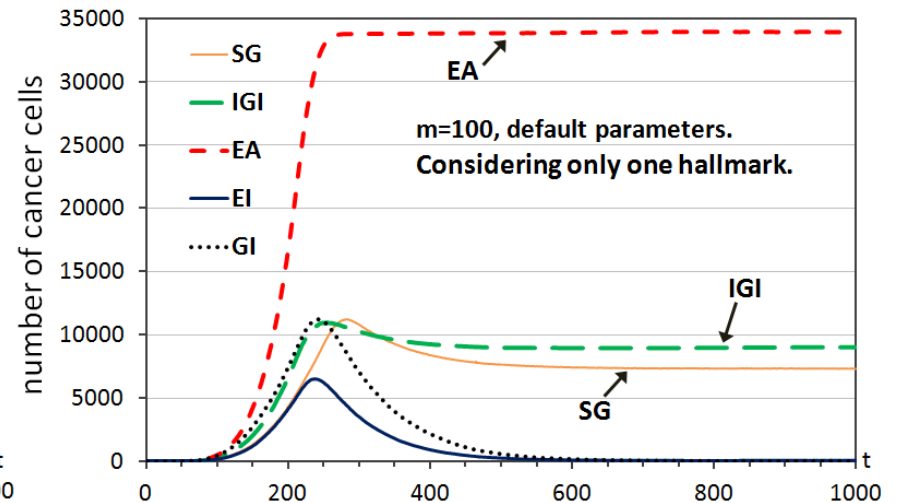
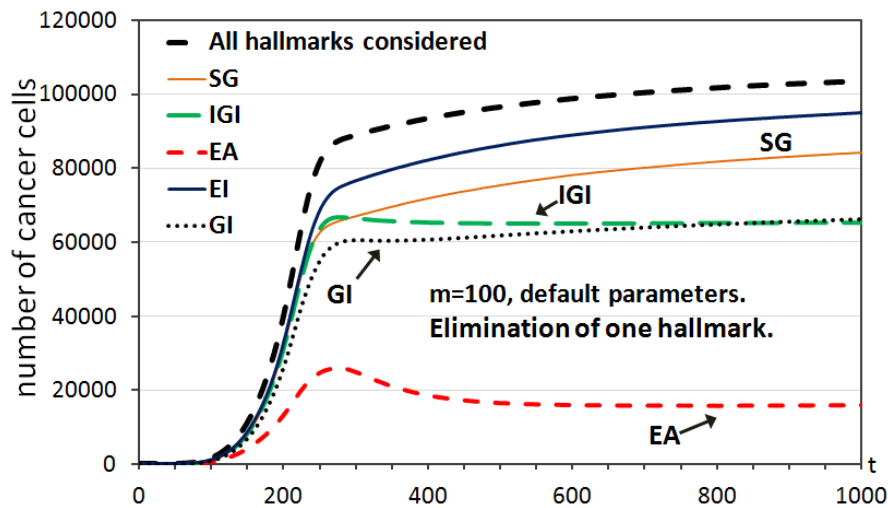


Snapshots of the cellular system at different time steps

Grid size= $10^6$ , mutation rate= $10^{-5}$

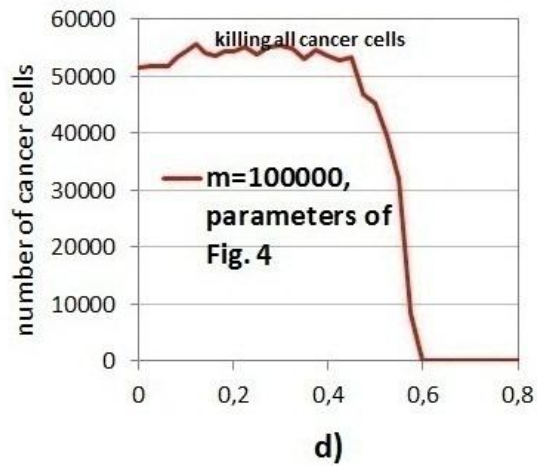
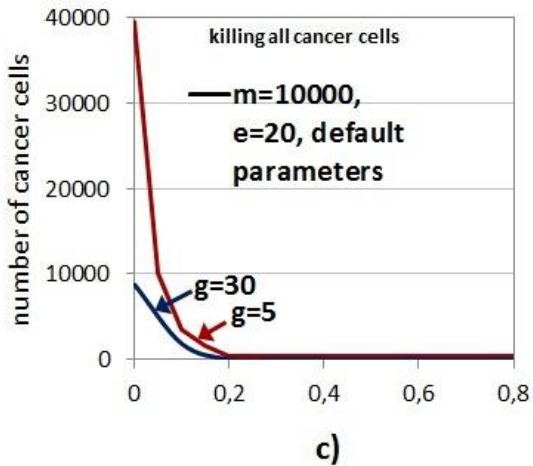
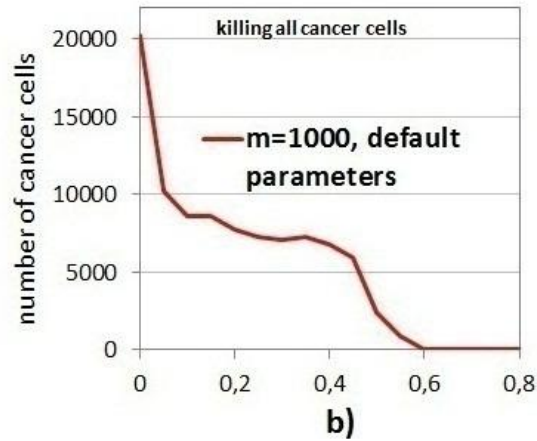
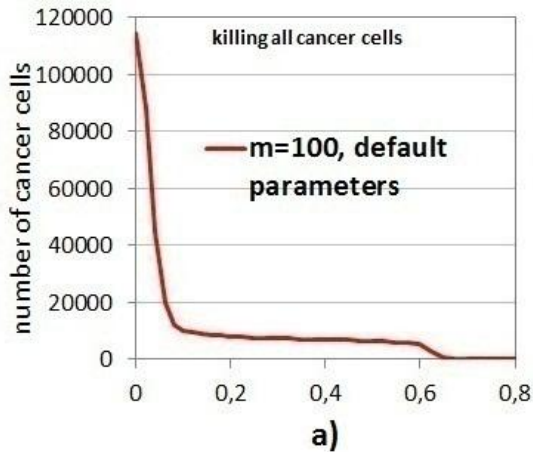
0

# Relevance of hallmarks

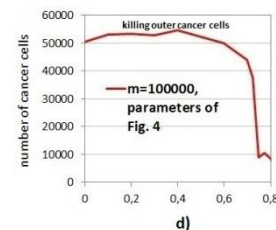
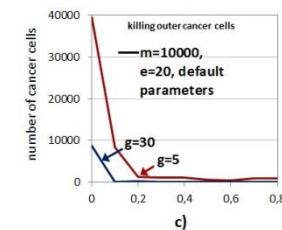
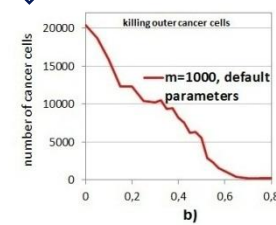
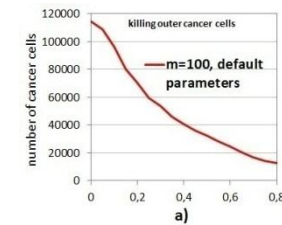


Effect of elimination of individual cancer hallmarks. Default parameters.

# Behavior transitions

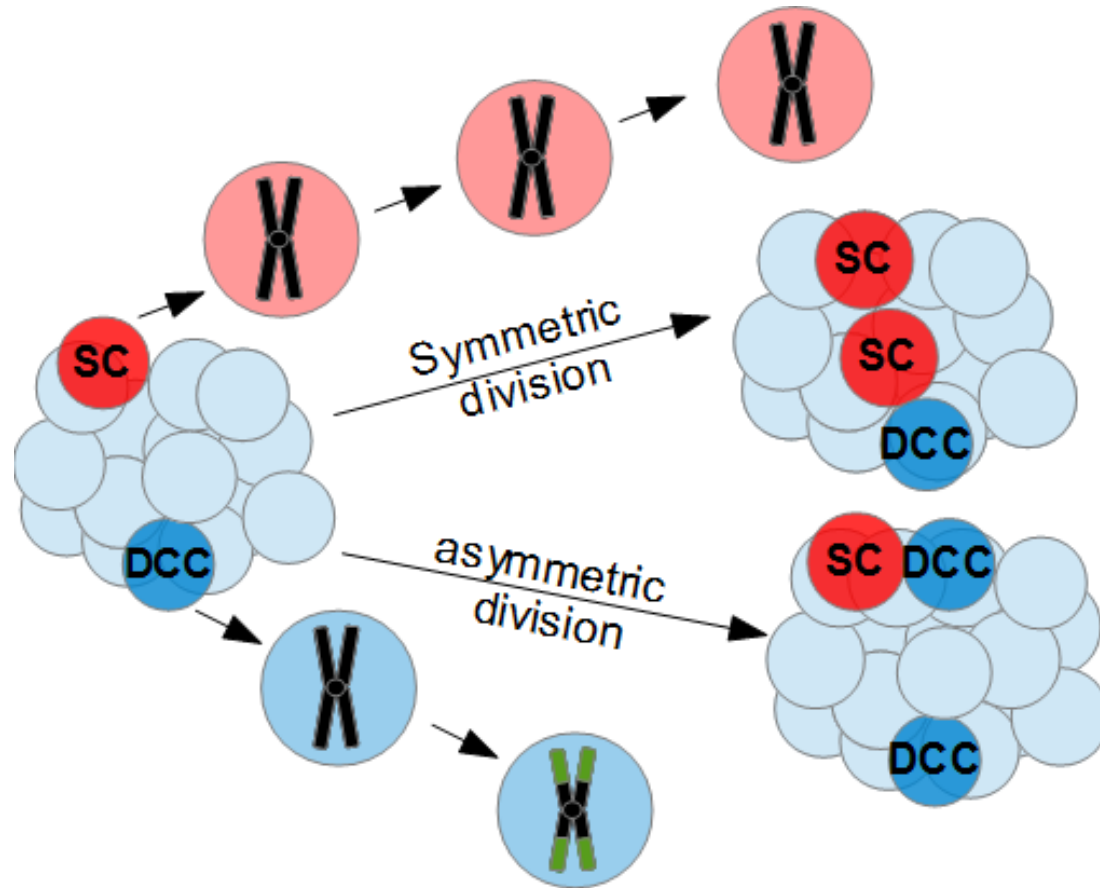


Killing only outer cancer cells



Effect of killing cancer cells during tumor growth for different killing probabilities and using four parameter sets.

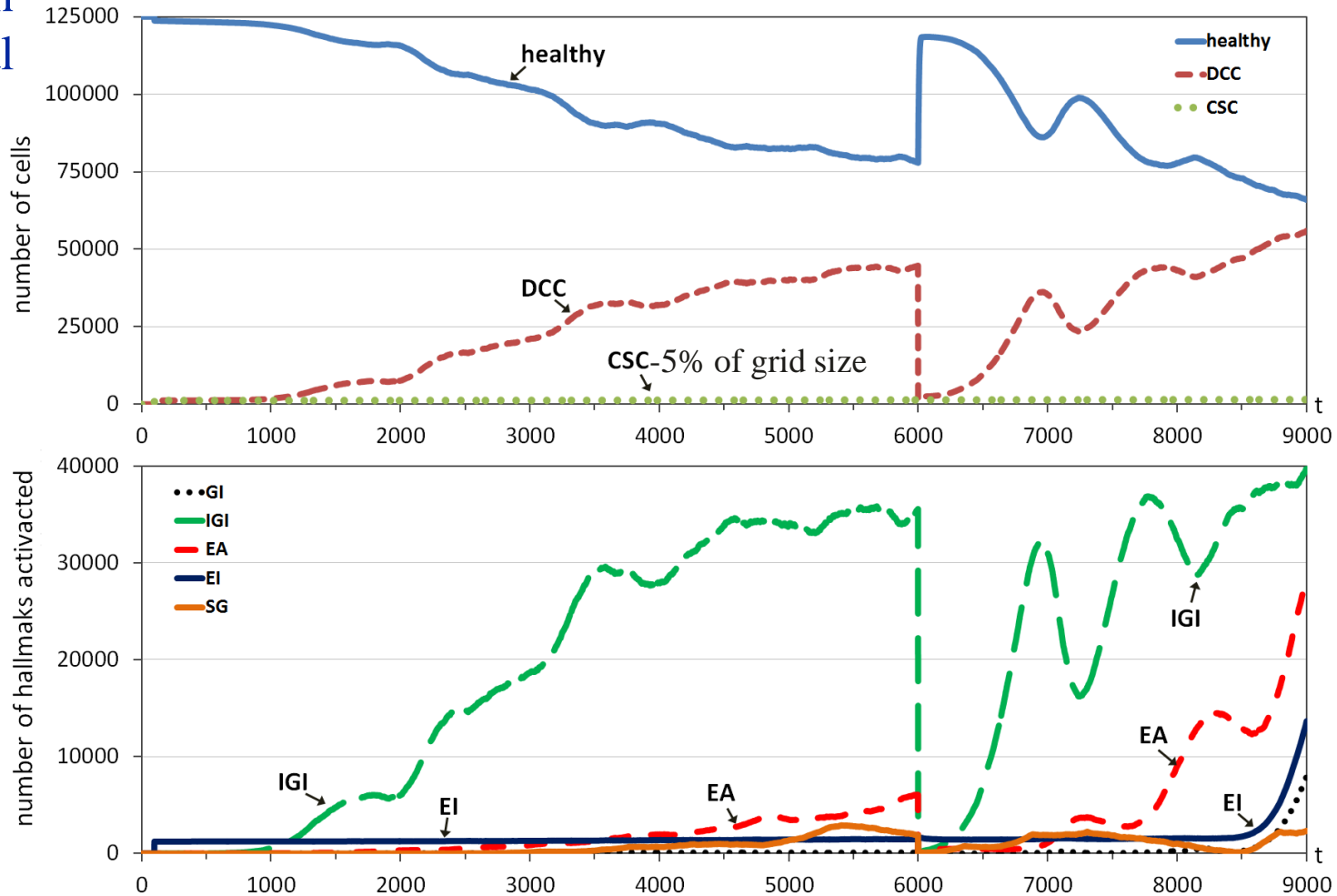
# Cancer Stem Cells



CSCs can divide symmetrically or asymmetrically to produce Differentiated Cancer Cells (DCCs) with limited proliferative capability.

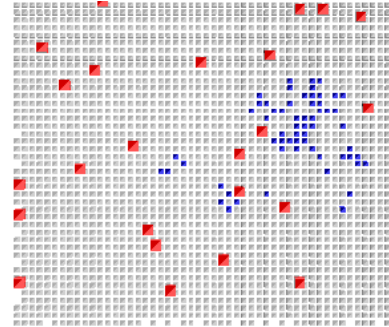
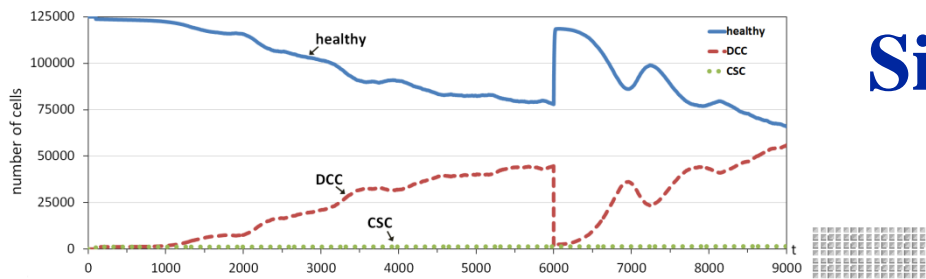
Example  
with high  
invasion  
potential

# Simulating Cancer Stem Cells

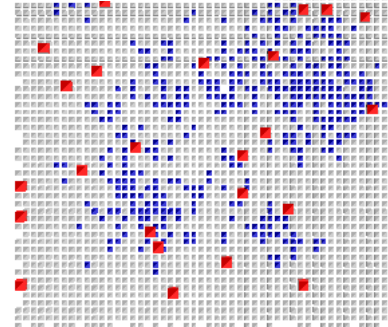


Introduction of Cancer Stem Cells (CSCs) in the multicellular system evolution.  
Standard parameters and  $g=5$ .

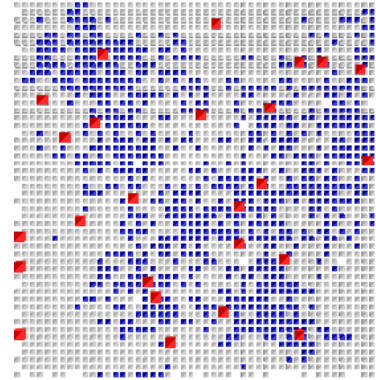
# Simulating Cancer Stem Cells



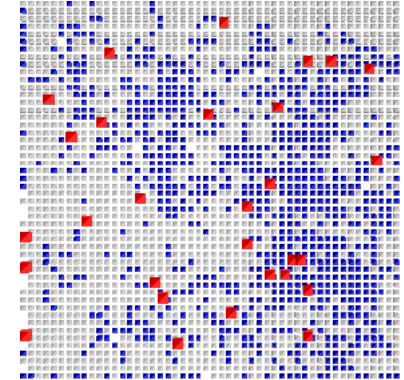
t=1500



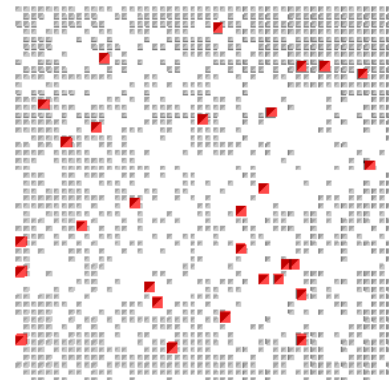
t=2500



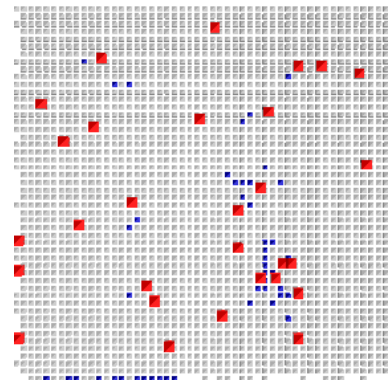
t=3500



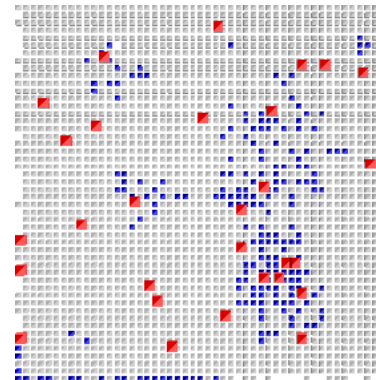
t=5500



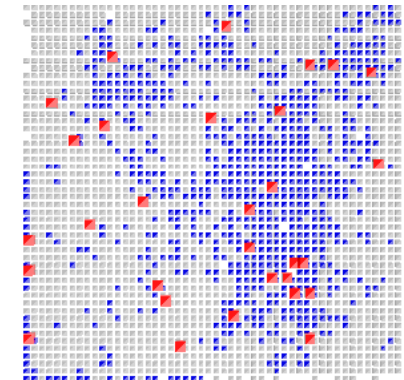
t=6000



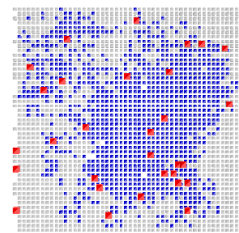
t=6250



t=6500



t=7000



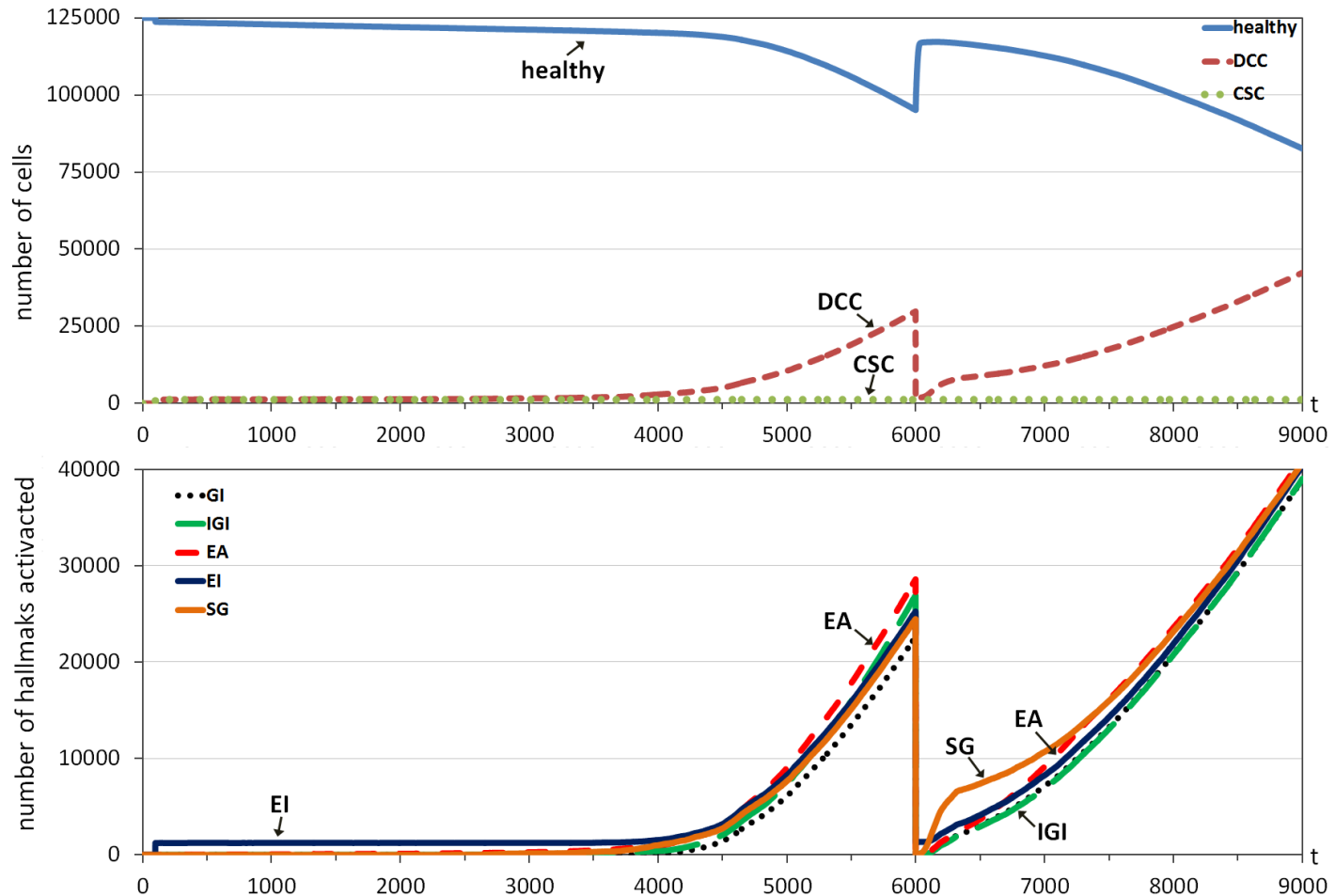
t=8500

Snapshots of 2D central sections

Gray: **Healthy** cells, Blue: **DCCs**, Red-enlarged size: **CSCs**.

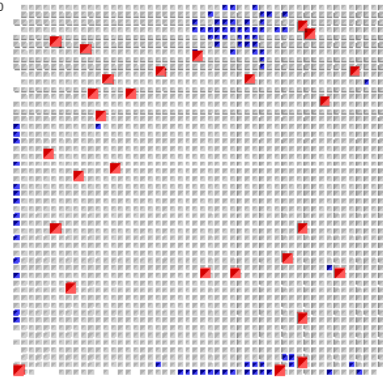
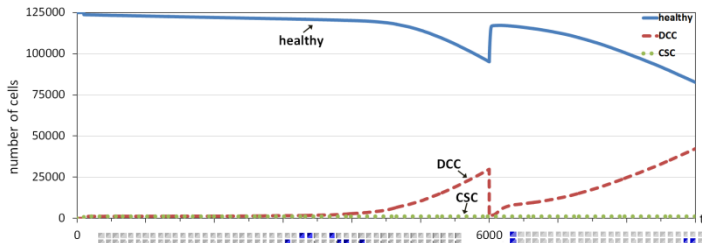
Example  
with high  
mutation  
rate

# Simulating Cancer Stem Cells

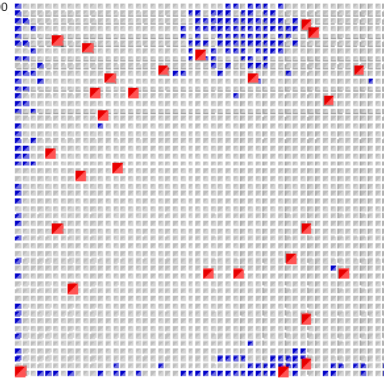


Standard parameters and  $m=1000$ .

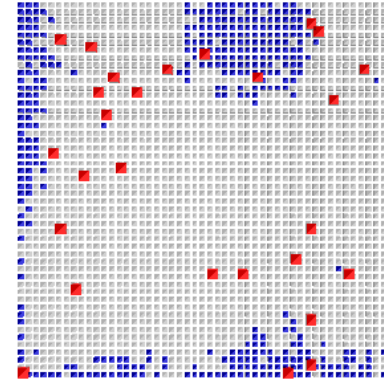
# Simulating Cancer Stem Cells



t=4500



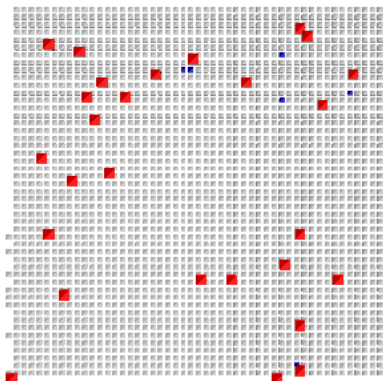
t=5000



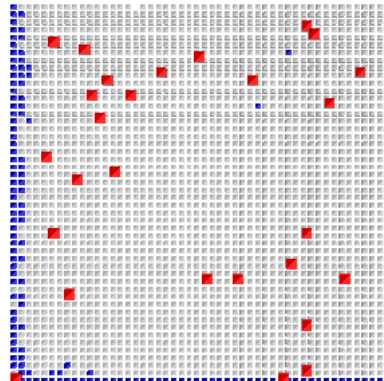
t=5500



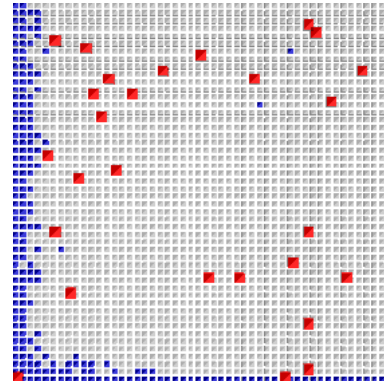
t=6000



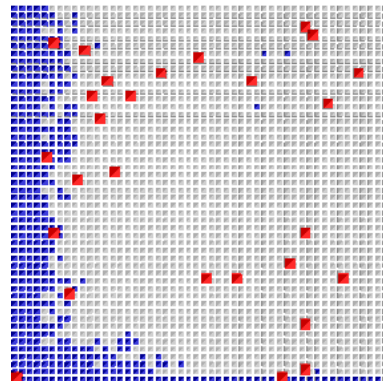
t=6100



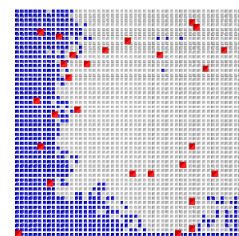
t=6500



t=7000



t=8000



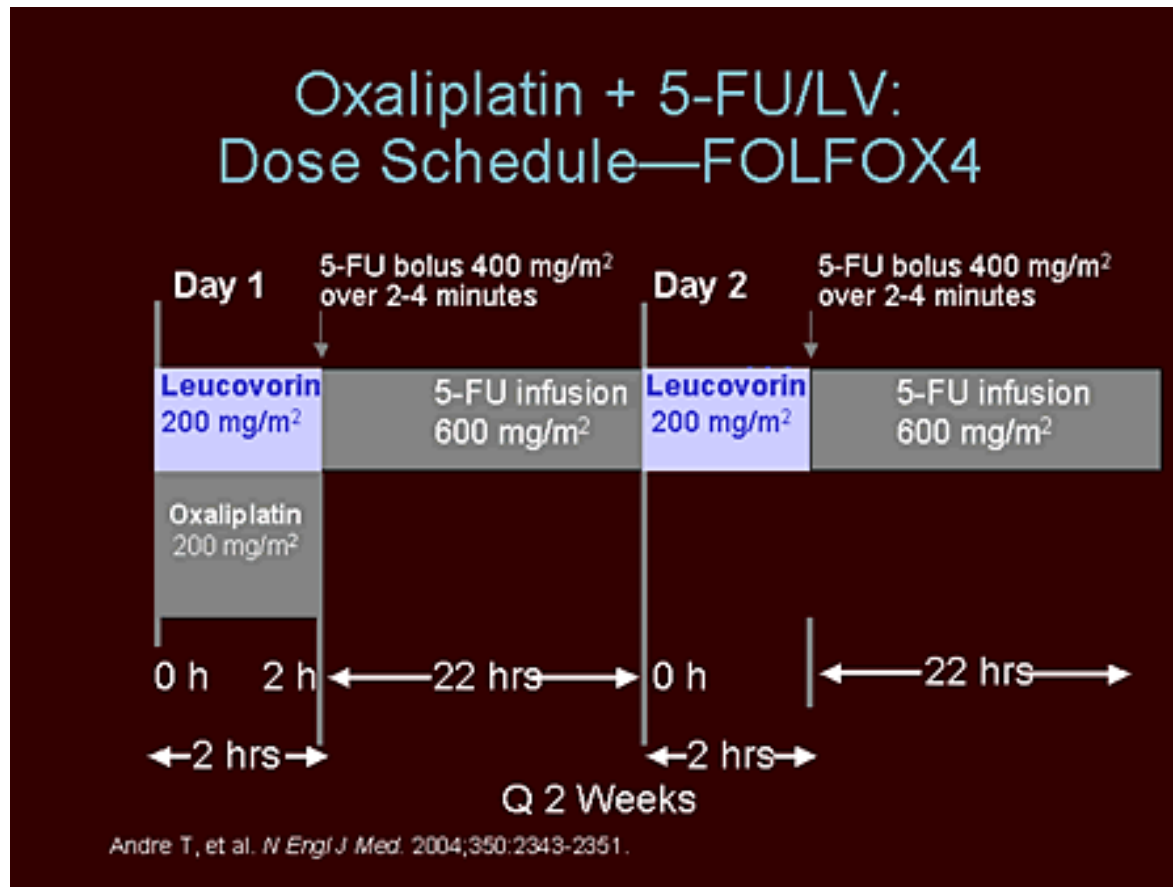
t=9000

Snapshots of 2D central sections

Gray: **Healthy** cells, Blue: **DCCs**, Red-enlarged size: **CSCs**.



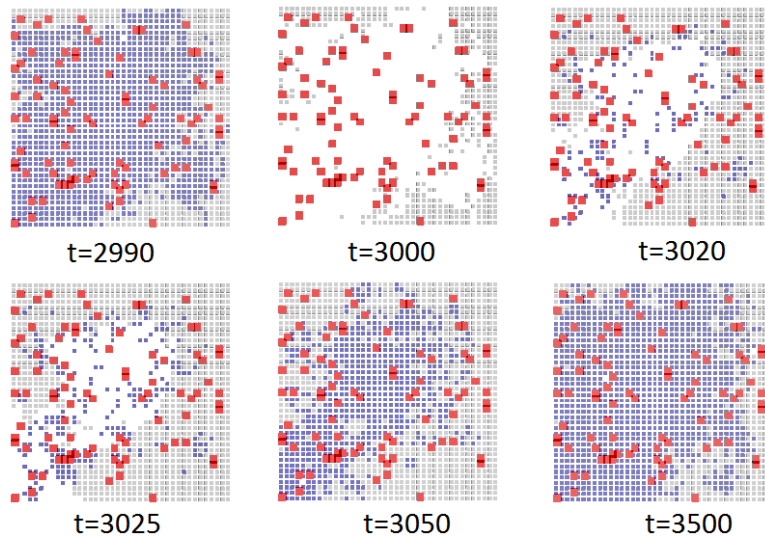
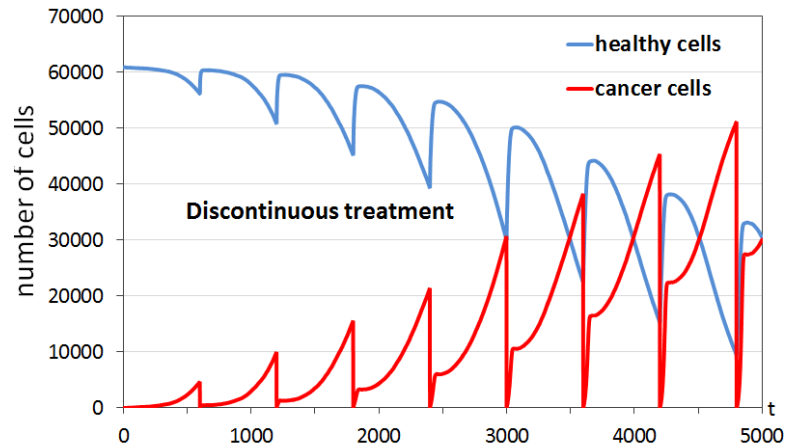
# Treatment options



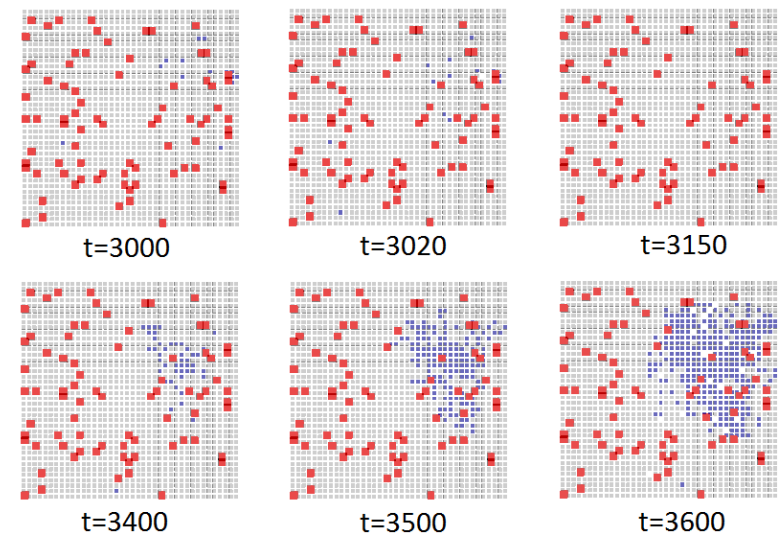
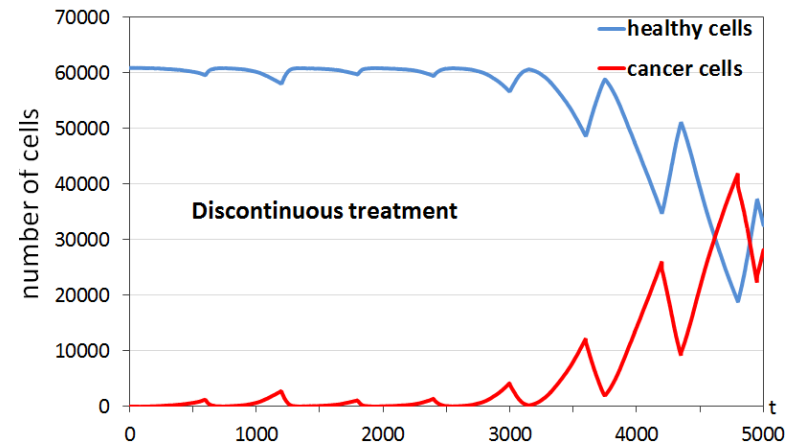
FOLFOX4 treatment for colon cancer

# Treatment options

Treatment applied every 600 time iterations killing the 100% of the DCCs in such iterations.



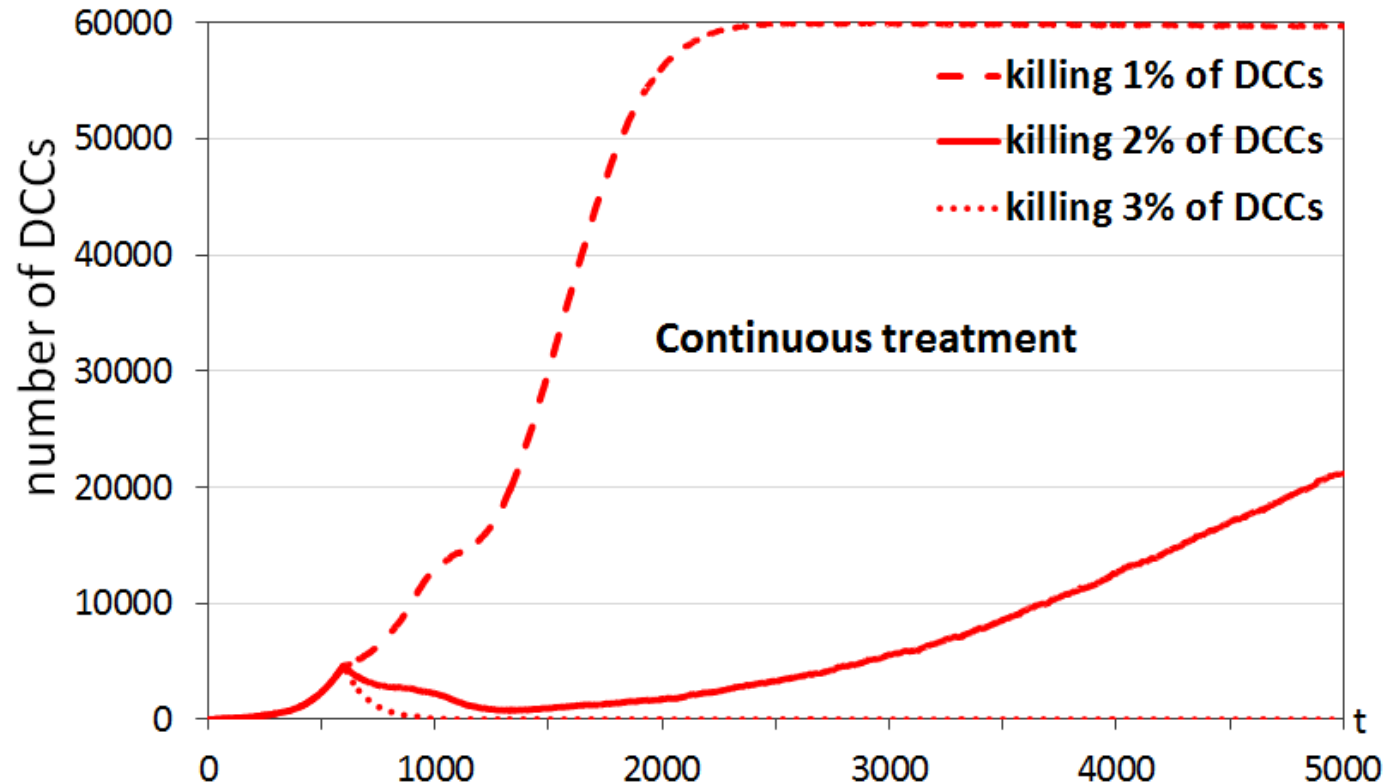
Treatment applied every 600 time iterations killing the 5% of the DCCs during the next 150 iterations.



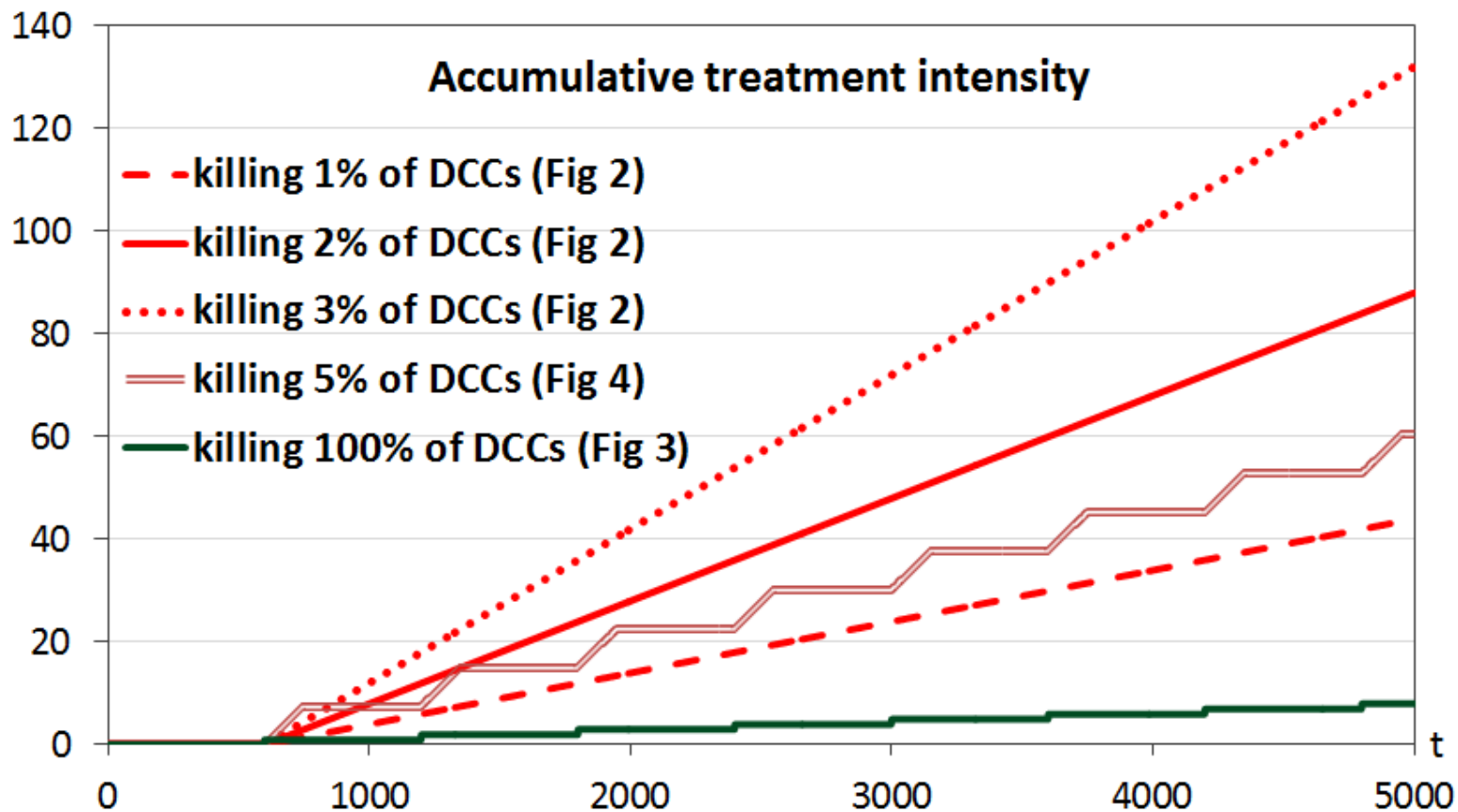
2D snapshots of the central part of the grid at given time iterations (Colors: Gray - healthy cells, Blue - DCCs, Red-enlarged size - CSCs).

# Treatment options

Continuous treatments that kill 1%, 2% and 3% of DCCs are applied in every time iteration.



# Treatment options



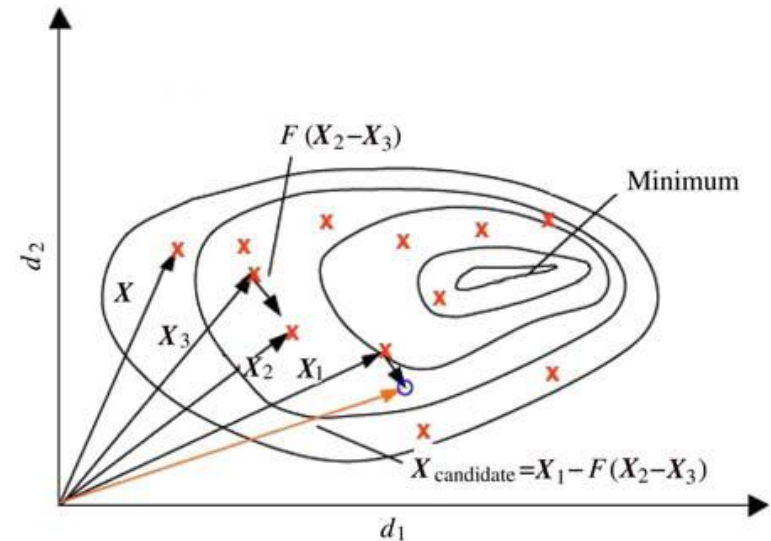
Accumulative treatment intensity across iterations

# Differential Evolution

In this application:  $F:0.9$

$CR:0.9$

$X_1$  selected with tournament



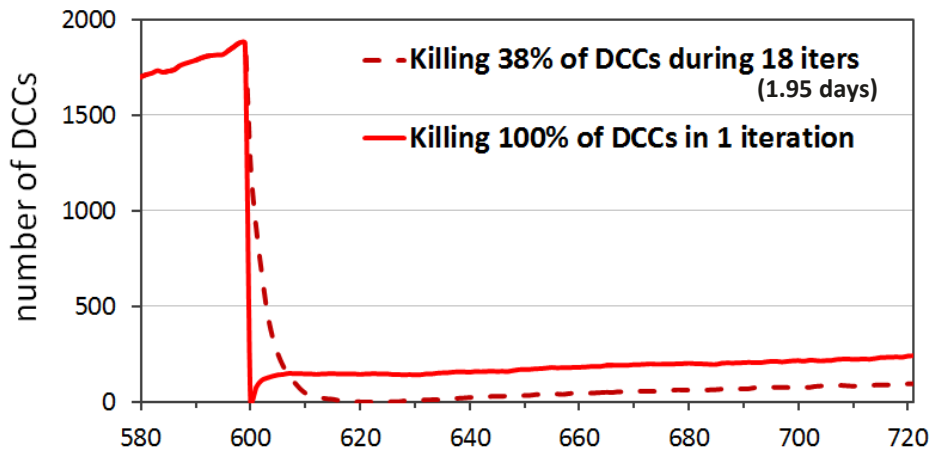
Encoded parameters: treatment intensity, duration and period.

50 generations, grid size of 64000 sites.

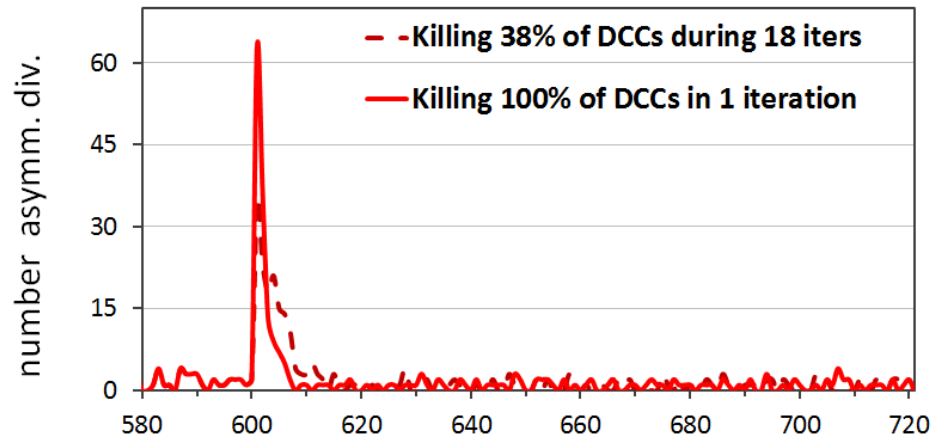
Hallmark parameters:  $g = 5$ ,  $m = 1000$ , others - standard values

Fitness: DCCs after a given number of iterations (100) once the treatment is stopped +  
Intensity used during the treatment application.

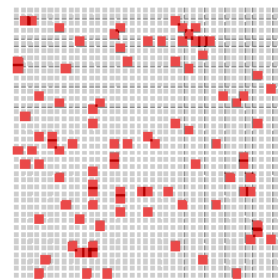
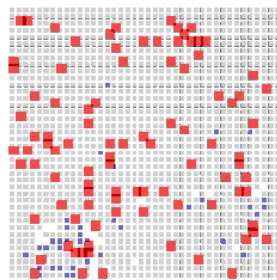
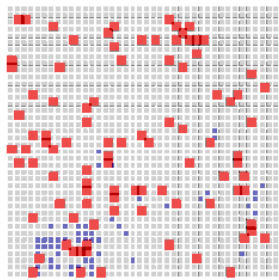
# Evolved treatment strategies



Evolution of DCCs

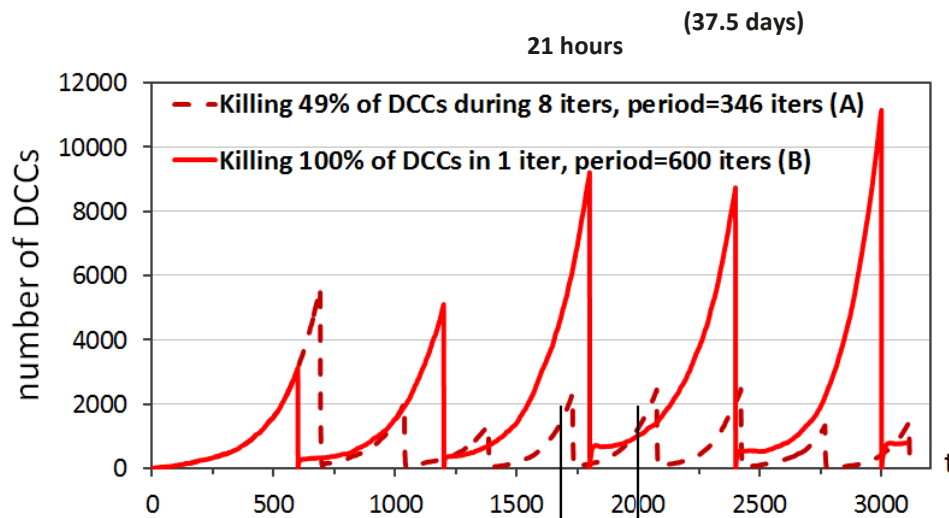


CSC asymmetric divisions

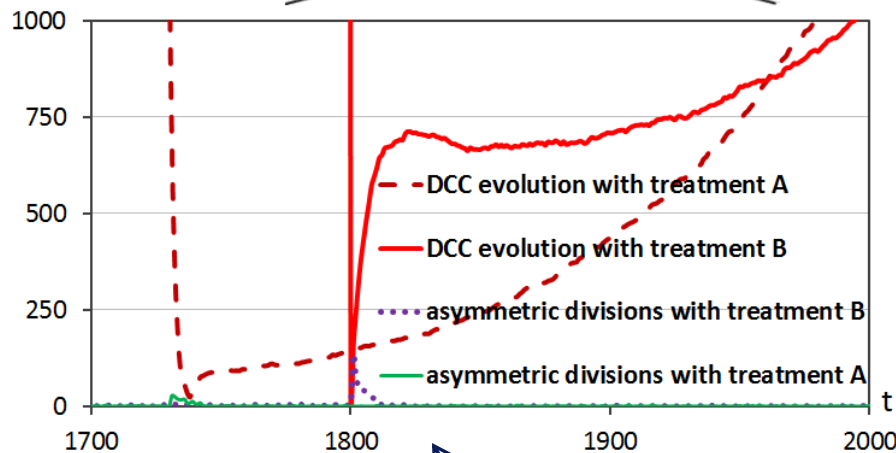


Treatment intensity and duration optimized. Treatments begin in time iteration  $t=600$ . The best evolved treatment kills 38% of DCCs in the next 18 time iterations. A high-intensity treatment that kills the 100% of DCCs only at  $t=600$  is included for comparison.

# Evolved treatment strategies



← Evolution of DCCs



← CSC asymmetric divisions

Treatment period, intensity and duration optimized. Treatments do not begin before time iteration  $t=600$ . The best evolved treatment kills 49% of DCCs during 8 time iterations and a period of 346 iterations (treatment A). A high-intensity treatment that kills the 100% of DCCs every 600 iterations is included for comparison (treatment B).

# Conclusions

The hallmark implications for cell population dynamics are difficult to foresee without a simulation model.

Our focus was on the dependences of the first phases of cancer growth on the hallmark parameters:

- In a CSC context, the model predicts and is in agreement with the clinical observations describing **increased growth speed and enhanced invasion in the relapsing malignancy**.
- Using EC: Treatments should be maintained during very few days, avoiding high intensities and, consequently, with longer periods than the ones used in standard treatments. The aim is to **make more difficult CSC proliferation** and consequently their differentiation to minimize their future effect on a possible tumor regrowth

The modeling can lead to a better understanding and characterization of the underlying biological processes involved.



- Adami, C. (1998), *Introduction to artificial life*, Telos-Springer Verlag.
- Ilachinski, A. (2001), *Cellular automata. A discrete universe*. World Scientific.
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